The relationship between T cell HLA-DR expression and intravenous immunoglobulin treatment response

in Kawasaki disease

Running title: HLA-DR and IVIG in Kawasaki disease

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Abstract

Background: Kawasaki disease (KD) is an early childhood febrile illness caused by vasculitis. Administration of intravenous immunoglobulin (IVIG) is now accepted as the standard treatment for KD. However, IVIG treatment is not effective in approximately 15% of KD children. Some reports have presented evidence of immunological responses in IVIG-resistant KD. We assessed the possibility that T cell activation was a contributory mechanism underlying this phenomenon.

Methods: We analyzed HLA-DR expression on peripheral blood CD4⁺ and CD8⁺ T cells in 82 children with KD who were admitted to our hospital between October 2007 and February 2012. We compared the percentages of HLA-DR⁺CD4⁺ T cells among CD4⁺ T cells and HLA-DR⁺CD8⁺ T cells among CD8⁺ T cells between the IVIG-effective and IVIG-resistant groups.

Results: Among 82 subjects, 51 children had IVIG-effective KD, and 31 children had IVIG-resistant KD. The percentages of HLA-DR⁺CD4⁺ among CD4⁺ T cells and HLA-DR⁺CD8⁺ T cells among CD8⁺ T cells in the IVIG-effective group were significantly lower than those in the IVIG-resistant group.

Conclusion: Our results suggest that decreased T cell HLA-DR expression is associated with IVIG-effective KD. It is likely that T cell HLA-DR expression is useful for predicting IVIG response in the acute phase of KD.

Keywords: Kawasaki disease, intravenous immunoglobulin, T cell, HLA-DR, CD4, CD8

Introduction

Kawasaki disease (KD) is an acute febrile illness associated with the development of vasculitis (1, 2). The most serious complication of KD is coronary arterial lesion (CAL) (3). High-dose intravenous immunoglobulin (IVIG) reduces the incidence of CAL in the majority of KD patients (4-6), but IVIG treatment is not effective in 13–21% of cases (7). The underlying cause of KD remains unknown, and many details of the immune response have not been characterized; however, it is known that all components of the immune system are involved (8). Few previous studies have addressed the role of different peripheral T cell populations in the acute phase of KD (9, 10). With respect to adaptive immunity, autopsy studies of children who died during the acute phase clearly indicate that CD4⁺ and CD8⁺ T cells participate in the transmural infiltration of the coronary arterial wall (11). HLA-DR is known as a marker of T cell activation (12-14); therefore, we investigated the relationship between HLA-DR expression on peripheral blood CD4⁺ and CD8⁺ T cells and IVIG response in children with KD.

Results

The parents of the patients enrolled in this study provided informed consent. This study included 82 children with KD who were admitted to the Department of Pediatrics at Yamaguchi University Hospital between October 2007 and February 2012. There were 54 boys and 28 girls, aged 2.8 ± 2.6 years (mean, 2.8 years; median, 1.8 years). All

the patients fulfilled the criteria described in the fifth revision of the Diagnostic Guidelines for Kawasaki Disease (15). We administered IVIG (2 g/kg) and oral aspirin (30 mg/kg daily) to all patients. We considered the cases of patients who showed reduced fever within 48 h after the end of IVIG infusion as IVIG-effective cases (Group A, 51 patients, 31 boys, 20 girls; aged 2.5 ± 2.3 years; mean, 2.5 years; median, 1.7 years), and those of patients with continued fever as IVIG-resistant cases (Group B, 31 patients, 23 boys, 8 girls; aged 3.8 ± 3.5 years; mean, 3.8 years; median, 3.1 years). Table 1 lists the clinical features of children in both groups.

The CD4⁺ T cell and HLA-DR⁺CD4⁺ T cell results are shown in Table 2 and Figure 1. The percentage of

HLA-DR⁺CD4⁺ T cells among peripheral blood mononuclear cells (PBMCs) was significantly lower in Group A than in Group B (p = 0.005). The HLA-DR⁺CD4⁺ T cell count and the percentage among CD4⁺ T cells in Group A were significantly lower than those in Group B (p = 0.004 and p = 0.003, respectively).

The CD8⁺ T cell and HLA-DR⁺CD8⁺ T cell results are shown in Table 3 and Figure 2. CD8⁺ T cells and HLA-DR⁺CD4⁺ T cells exhibited very similar trends to those observed for CD4⁺ T cells and HLA-DR⁺CD4⁺ T cells. The count of HLA-DR⁺CD8⁺ T cells was significantly lower in Group A than in Group B (p = 0.019). The percentage of HLA-DR⁺CD8⁺ T cells among CD8⁺ T cells in Group A was significantly less than that in Group B (p = 0.046). The relationship of the percentage of HLA-DR⁺CD8⁺ T cells among CD8⁺ T cells among CD

Discussion

We previously reported on the pathogenesis of KD (16-22, 25, 26). In terms of changes in peripheral blood immunocyte counts in KD patients, increases in white blood cell number correlated with increases in neutrophil count, but there was no great variation in mononuclear cell count (16). However, the numbers of CD14⁺ monocytes/macrophages and CD19⁺ B cells was increased among mononuclear cells, whereas those of CD4⁺ and CD8⁺ T cells were slightly decreased (16, 17). It has been reported that activation of nuclear factor- κ B (NF- κ B) is evident in peripheral blood CD14⁺ monocytes/macrophages, and this activation is reduced by IVIG (18-20). The number of peripheral blood CD14⁺CD16⁺ (Fc γ RIII) monocytes/macrophages is also increased in KD, and IVIG strongly suppresses Fc γ RIII expression in monocytes/macrophages (21-22). Many studies have demonstrated the effect of IVIG treatment on KD, but few of these have utilized flow cytometry (23, 24). In this study, we investigated the relationship between HLA-DR expression on peripheral blood CD4⁺ and CD8⁺ T cells and IVIG responsiveness.

We previously reported that HLA-DR expression is low in $CD4^+$ and $CD8^+$ T cells during the acute phase of KD (17, 25, 26), and another group corroborated the observation that HLA-DR⁺CD8⁺ T cell number does not increase during the acute phase of KD (27). An additional study showed that soluble interleukin-2 receptor (a marker of T cell activation) is elevated in IVIG-resistant KD (28). Here, we determined that HLA-DR expression levels on $CD4^+$ and $CD8^+$ T cells are significantly suppressed in IVIG-effective KD patients compared to IVIG-resistant

KD patients. These results support those of previous studies (17, 25, 26) and suggest that both $CD4^+$ and $CD8^+$ T cell activation is generally suppressed during the acute phase of KD. Moreover, they imply that patients with weak suppression of $CD4^+$ and $CD8^+$ T cell activation may not respond to high-dose IVIG therapy. Finally, these results indicate that HLA-DR expression on $CD4^+$ and $CD8^+$ T cells may be used as a predictive marker for the effectiveness of IVIG treatment in KD patients.

Methods

Flow cytometry

We collected heparinized whole blood from children in the acute phase of KD just before IVIG infusion. They were labeled with peridinin chlorophyll protein (PerCP)-conjugated anti-HLA-DR, FITC-conjugated mouse anti-human CD4, or FITC-conjugated mouse anti-human CD8. The samples were also labeled with FITC-conjugated mouse IgG1 and PerCP-conjugated mouse IgG2a as a negative control. Erythrocytes were lysed with fluorescence-activated cell sorting (FACS) Lysing Solution. The cells were centrifuged and resuspended in FACS Flow. For each sample, 5,000 cells were analyzed by a FACScaliber flow cytometer equipped with CellQuest software. All the reagents and equipment used for FACS analysis was purchased from BD Biosciences (Franklin Lakes, NJ, USA).

Statistical analysis

Differences between the two groups were analyzed using Mann-Whitney U tests, Fisher exact test, and *p* values less than 0.05 were considered statistically significant. The analyses and calculations were performed using the Statistical Package for Social Sciences (SPSS), version 12.0 (SPSS Inc., Chicago, IL, USA).

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References

- Kawasaki T, Kosaki G, Osawa S, Shigematsu I, Yanagawa S. A new infantile acute febrile mucocutaneous lymph node syndrome (MLNS) prevailing in Japan. Pediatrics 1974;54:271–6.
- 2. Fujiwara H, Hamashima Y. Pathology of the heart in Kawasaki disease. Pediatrics 1978;61:100-7.
- Kato H, Sugimura T, Akagi T, et al. Long-term consequences of Kawasaki disease. A 10- to 21-year follow-up study of 594 patients. Circulation 1996;94:1379–85.
- Furusho K, Kamiya T, Nakano H, et al. High-dose intravenous gammaglobulin for Kawasaki disease. Lancet 1984;2:1055–8.
- Newburger JW, Takahashi M, Burns JC, *et al.* The treatment of Kawasaki syndrome with intravenous gamma globulin. N Engl J Med 1986;315:341–7.
- 6. Newburger JW, Takahashi M, Beiser AS, *et al*. A single intravenous infusion of gamma globulin as compared with four infusions in the treatment of acute Kawasaki syndrome. N Engl J Med 1991;324:1633–9.
- Sleeper LA, Minich LL, McCrindle BM, *et al.* Evaluation of Kawasaki disease risk- scoring systems for intravenous immunoglobulin resistance. J Pediatr 2011;158:831–5.
- 8. Burns JC. The riddle of Kawasaki disease. N Engl J Med 2007;356:659-61.
- Franco A, Shimizu C, Tremoulet AH, Burns JC. Memory T-cells and characterization of peripheral T-cell clones in acute Kawasaki disease. Autoimmunity 2010;43:317–24.

- Brogan PA, Shah V, Clarke LA, Dillon MJ, Klein N. T cell activation profiles in Kawasaki syndrome. Clin Exp Immunol 2008;151:267–74.
- 11. Brown TJ, Crawford SE, Cornwall ML, Garcia F, Shulman ST, Rowley AH. CD8 T lymphocytes and macrophages infiltrate coronary artery aneurysms in acute Kawasaki disease. J Infect Dis 2001; 184:940–3.
- 12. Reddy M, Eirikis E, Davis C, Davis HM, Prabhakar U. Comparative analysis of lymphocyte activation marker expression and cytokine secretion profile in stimulated human peripheral blood mononuclear cell cultures: an in vitro model to monitor cellular immune function. J Immunol Methods 2004;293:127–42.
- 13. Starska K, Glowacka E, Kulig A, Lewy-Trenda I, Brys M, Lewkowicz P. The role of tumor cells in the modification of T lymphocytes activity the expression of the early CD69(+), CD71(+) and the late CD25(+), CD26(+), HLA/DR(+) activation markers on T CD4(+) and CD8(+) cells in squamous cell laryngeal carcinoma. Part I. Folia Histochem Cytobiol 2011;49:579–92.
- Tincati C, Bellistri GM, Ancona G, Merlini E, d'Arminio Monforte A, Marchetti G. Role of in vitro stimulation with lipopolysaccharide on T-cell activation in HIV-infected antiretroviral-treated patients. Clin Dev Immunol 2012;2012:935425.
- The Japan Kawasaki Disease Research Committee. Diagnostic guidelines of Kawasaki disease. 5th revised
 2002. http://www.kawasaki-disease.org/ diagnostic/index.html. Accessed 3 Oct 2008.
- 16. Furukawa S, Matsubara T, Yabuta K. Mononuclear cells subsets and coronary artery lesions in Kawasaki

disease. Arch Dis Child 1992;67:706-8.

- 17. Matsubara T, Ichiyama T, Furukawa S. Immunological profile of peripheral blood lymphocytes and monocytes/macrophages in Kawasaki disease. Clin Exp Immunol 2005;141:381–7.
- Ichiyama T, Yoshitomi T, Nishikawa M, *et al*. NF-κB activation in peripheral blood monocytes/macrophages and T cells during acute Kawasaki disease. Clin Immunol 2001;99:373–7.
- Ichiyama T, Ueno Y, Hasegawa M, Niimi A, Matsubara T, Furukawa S. Intravenous immunoglobulin inhibits NF-kappaB activation and affects Fcgamma receptor expression in monocytes/macrophages. Naunyn Schmiedebergs Arch Pharmacol 2004;369:428–33.
- 20. Ichiyama T, Ueno Y, Isumi H, Niimi A, Matsubara T, Furukawa S. An immunoglobulin agent (IVIG) inhibits NF-kappaB activation in cultured endothelial cells of coronary arteries in vitro. Inflamm Res 2004;53:253–6.
- 21. Katayama K, Matsubara T, Fujiwara M, Koga M, Furukawa S. CD14⁺CD16⁺ monocyte subpopulation in Kawasaki disease. Clin Exp Immunol 2000;121:566–70.
- 22. Ichiyama T, Ueno Y, Hasegawa M, Ishikawa Y, Matsubatra T, Furukawa S. Intravenous immunoglobulin does not increase Fc γ RIIB expression on monocytes/macrophages during acute Kawasaki disease. Rheumatology 2005;44:314–7.
- 23. Abe J, Ebata R, Jibiki T, Yasukawa K, Saito H, Terai M. Elevated granulocyte colony-stimulating factor levels predict treatment failure in patients with Kawasaki disease. J Allergy Clin Immunol 2008;122:1008–13.

- 24. Jia S, Li C, Wang G, Yang J, Zu Y. The T helper type 17/regulatory T cell imbalance in patients with acute Kawasaki disease. Clin Exp Immunol 2010;162:131–7.
- 25. Furukawa S, Matsubara T, Tsuji K, Motohashi T, Okumura K, Yabuta K. Serum soluble CD4 and CD8 levels

in Kawasaki disease. Clin Exp Immunol 1991;86:134-9.

- 26. Matsubara T, Katayama K, Matsuoka T, Fujiwara M, Koga M, Furukawa S. Decreased interferon-gamma (IFN-gamma)-producing T cells in patients with acute Kawasaki disease. Clin Exp Immunol 1999;116:554–7.
- 27. Ehara H, Kiyohara K, Izumisawa Y, Ito T. Early activation does not translate into effector differentiation of peripheral CD8T cells during the acute phase of Kawasaki disease. Cell Immunol 2010;265:57–64.
- 28. Suzuki H, Suenaga T, Takeuchi T, Shibuta S, Yoshikawa N. Marker of T-cell activation is elevated in

refractory Kawasaki disease. Pediatr Int 2010;52:785-9.

Figure legends

Figure 1. The relationship between IVIG effectiveness and HLA-DR⁺CD4⁺ T cells. The percentages of peripheral blood HLA-DR⁺CD4⁺ T cells among CD4⁺ T cells in patients with KD were analysed by flow cytometry. Group A: IVIG-effective KD, Group B: IVIG-resistant KD. ** p < 0.01. Horizontal lines indicate means.

Figure 2. The relationship between IVIG effectiveness and HLA- DR^+CD8^+ T cells. The percentages of peripheral blood HLA- DR^+CD8^+ T cells among $CD8^+$ T cells in patients with KD were analysed by flow cytometry. Group A: IVIG-effective KD, Group B: IVIG-resistant KD. * p < 0.05. Horizontal lines indicate means.

	KD (r			
-	IVIG effective (n = 51)	IVIG resistant (n = 31) —	P value	
	(II - 31) Group A	Group B	A vs. B	
Median age (years) (range)	2.5 (1.0-7.5)	3.8 (0.2–9.1)	0.257	
Male : Female	31:20	23:8	0.240†	
Median starting days of IVIG (range)	5 (3-9)	4 (3-11)	0.317	
Coronary arterial leision (n)	1	7	0.004**†	

** *P* < 0.01.

[†] Fisher exact test was used

	KD (n = 82)		
	IVIG effective (n = 51) Group A	IVIG resistant (n = 31) Group B	P value A vs. B
PBMCs (/µl)	4,318 ± 1,829	5,499 ± 4,570	0.339
CD4 ⁺ T cells among PBMCs (%)	35.6 ± 10.7	34.4 ± 7.3	0.916
CD4 ⁺ T cells (/µl)	1,601 ± 957	1,876 ± 1,323	0.293
HLA-DR ⁺ CD4 ⁺ T cells among PBMCs (%)	1.7 ± 1.1	2.3 ± 1.0	0.005**
HLA-DR ⁺ CD4 ⁺ T cells (/µl)	70 ± 47	131 ± 110	0.004**
HLA-DR ⁺ CD4 ⁺ T cells among CD4 ⁺ T cells (%)	5.1 ± 3.4	6.8 ± 2.8	0.003**

** *P* < 0.01.

	KD (n = 82)		
	IVIG effective (n = 51)	IVIG resistant (n = 21)	P value
	(II – 51) Group A	(n = 31) - Group B	A vs. B
PBMCs (/µl)	4,318 ± 1,829	5,499 ± 4,570	0.339
CD8 ⁺ T cells among PBMCs (%)	23.8 ± 7.8	23.8 ± 6.7	0.863
CD8 ⁺ T cells (/µl)	$1,000 \pm 493$	1,315 ± 1,116	0.202
HLA-DR ⁺ CD8 ⁺ T cells among PBMCs (%)	2.5 ± 2.2	3.4 ± 2.9	0.071
HLA-DR ⁺ CD8 ⁺ T cells (/µl)	96 ± 81	186 ± 192	0.019*
HLA-DR ⁺ CD8 ⁺ T cells among CD8 ⁺ T cells (%)	9.9 ± 7.4	13.7 ± 10.1	0.046*

* *P* < 0.05.







HLA-DR⁺CD8⁺ T cells among CD8⁺ T cells (%)

Figure 2