Anti-ganglioside Autoantibody in a Regional Variant of Guillain-Barré Syndrome

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Guillain-Barre syndrome (GBS) is an immune-mediated peripheral polyneuropathy characterized by acute onset of symmetrical limb weakness and areflexia. Patients with typical GBS have greater *leg than arm* weakness with "ascending" progression. Some, however, have muscle weakness only at the oropharynx, neck, and proximal upper limb muscles, a "descending" pattern of weakness appearing as the illness progresses. Ropper¹⁾ proposed that such patients had a variant of GBS, "pharyngeal-cervical-brachial weakness (PCB)". *Elsewhere* we confirmed by statistical analysis that PCB is a distinct clinical variant of GBS².

Gangliosides, important constituents of the plasma membrane, comprise a family of sialic acid-containing glycolipids that are concentrated mainly in the nervous system. In the last decade, autoantibodies against gangliosides have been shown to increase in sera during the acute phase of GBS. The presence of anti-ganglioside antibody is closely related to certain GBS variants. Serum IgG anti-GQ1b ganglioside antibody often is present in Miller Fisher syndrome (MFS)³⁾, a variant of GBS with the triad of ophthalmoplegia, ataxia, and areflexia. IgG anti-GM1 ganglioside antibody is present in the pure motor variant⁴⁾. A patient with PCB-like symptoms who had anti-GT1a and anti-GD1a IgG antibodies has been reported⁵⁾, but which antibody is associated with this PCB variant was not clarified. We assumed that the former antibody is specific for PCB because the latter also is detected in the pure motor variant $^{6)}. \label{eq:scalar}$

We made a prospective search of PCB patients and found 1 with PCB⁷). As expected, this patient had high IgG anti-GT1a antibody titer, as did most patients with MFS. An absorption study, however, showed that the IgG anti-GT1a antibody in the PCB patient was not absorbed by GQ1b ganglioside, whereas the antibody in the MFS patients was. The frequency of positive IgG anti-GT1a antibody did not differ in patients with and without bulbar palsy, a cardinal sign of PCB.

These findings indicate that IgG anti-GT1a antibodies, which do not cross-react with GQ1b, are specific to PCB and provide a diagnostic marker for it. It is important to differentiate PCB from botulism and myasthenia gravis, which have similar clinical features, as plasmapheresis is required treatment for PCB patients. Because IgG anti-GM1b antibody also is present in some PCB patients⁸⁾, we should routinely test for serum anti-GT1a and anti-GM1b IgG antibodies in order to select the most efficacious treatment.

GBS is considered a postinfectious disease, and *Campylobacter jejuni*, a leading cause of acute diarrhea, is an antecedent infectious agent in one-third of GBS patients⁹⁾. There is molecular mimicry between the lipopolysaccharides of *C*. *jejuni* and gangliosides of human nerve tissue¹⁰⁾, and its existence has been hypothesized as being a critical factor in the onset of GBS. GT1a- and GM1b-like structures are present in both human peripheral nerve and certain strains of C. *jejuni*. Interestingly, most of our PCB patients had a history of gastrointestinal illness before the onset of neurological symptoms. I speculate that the molecular mimicry between C. *jejuni* lipopolysaccharides and nerve tissue gangliosides functions in the induction of anti-GT1a and anti-GM1b IgG antibodies, thereby causing muscle weakness in the region where these epitopes are concentrated. It is necessary to determine the distribution of GT1a and GM1b gangliosides in the human nervous system.

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