A Versatile Petri Net Based Architecture for Modeling and Simulation of Complex Biological Processes

	Masao Nagasaki 1	${f Atsushi}$ Doi 2
	masao@ims.u-tokyo.ac.jp	atsushi@ib.sci.yamaguchi-u.ac.jp
	$Hiroshi Matsuno^3$	${f Satoru} \ {f Miyano}^1$
	matsuno@sci.yamaguchi-u.a	ac.jp miyano@ims.u-tokyo.ac.jp
Hu	man Genome Center, Institute	of Medical Science, University of Tokyo, 4-6-1
Shi	rokane-dai, Minato-ku, Tokvo 10	8-8639. Japan

² Graduate School of Science and Engineering, Yamaguchi University, 1677-1 Yoshida, Yamaguchi 753-8512, Japan

³ Faculty of Science, Yamaguchi University, Yoshida, Yamaguchi 753-8512, Japan

Keywords: biopathway, hybrid functional Petri net with extension, biological process modeling, simulation, p53, frameshift, alternative splicing, Huntington's Disease

Abstract

The research on modeling and simulation of complex biological systems is getting more important in Systems Biology. In this respect, we have developed Hybrid Function Petri net (HFPN) that was newly developed from existing Petri net because of their intuitive graphical representation and their capabilities for mathematical analyses. However, in the process of modeling metabolic, gene regulatory or signal transduction pathways with the architecture, we have realized three extensions of HFPN, (i) an entity should be extended to contain more than one value, (ii) an entity should be extended to handle other primitive types, *e.g.* boolean, string, (iii) an entity should be extended to handle more advanced type called object that consists of variables and methods, are necessary for modeling biological systems with Petri net based architecture. To deal with it, we define a new enhanced Petri net called hybrid functional Petri net with extension (HFPNe). To demonstrate the effectiveness of the enhancements, we model and simulate with HFPNe four biological processes that are difficult to represent with the previous architecture HFPN.

1 Introduction

1

Our knowledge about cellular processes with biological experiments is increasing at a rapidly growing pace. With the knowledge, one of the most important topics in the Systems Biology is to understand and predict a cell's behavior. Related to this, particularly needed is a software environment with which biological and medical scientists (users) can comfortably model and simulate biological processes in the cell. Especially among biological processes, modelings and simulation of gene regulations, metabolic pathways, and signal-transduction cascades (say biopathways) are usually focused on the targets of software platforms.

First of all, to model and simulate biopathways, architecture is necessary. Thus, in 1999, we surveyed which architecture is suitable when modeling and simulating biopathway for biological and medical scientists. At that time, there were ODE-based attempts for modeling and simulating chemical reactions - *e.g.* Gepasi [26], E-Cell [35] - and others - *e.g.* the Lisp [25] based architecture, QSIM [17] and our other work, the π -Calculus based architecture, Bio-Calculus [28]. Unfortunately, applications based on these architectures are not acceptable in their fields. This is due to poor GUI interfaces, *e.g.* lacking biopathway editors, or their architectures themselves. To overcome this situation, we came

to the conclusion that an architecture based on Petri nets should be suitable because of their intuitive graphical representation and their capabilities for mathematical analyses [29, 30, 31]. Then we have newly developed HFPN for easy biopathway modeling [22]. With HFPN, we can model rule based biological processes in biopathways, *e.g.* gene regulation and also ODEs-based kinetics, *e.g.* chemical processes in biopathways. In fact, with HFPN, we have modeled and simulated: the glycolytic pathway of *Escherichia coli*, gene regulation of circadian rhythms in *Drosophila*, boundary formation by notch signaling in *Drosophila* [23], and apoptosis induced by *Fas* ligand [21].

However, when modeling biopathways with HFPN, we have realized that three extensions will be useful for modeling and simulating more complicated biopathway processes (e.g. activities of enzymesfor a multi-modification protein,) and other biological processes that are not normally treated in biopathways (e.q. alternative splicings, frameshiftings). The first is, an entity should be extended to contain more than one value, such as list and pair, because, (i) every part in a cell should contain 3D information, e.g. position and speed, (ii) proteins often have many modified states, e.g. p53 has known sixteen phosphorylation positions and two acetylation positions as in Table 8 and modified states of p53 can be 2^{18} [16]. In HFPN, an entity can contain only one value. The second is, HFPN should be extended to handle other primitive types, e.g. boolean, string, because major parts in a cell contain information similar to strings such as DNA sequences, mRNA sequences and protein sequences. In HFPN, an entity has only two types, discrete (non-negative integers) and continuous (non-negative real numbers). The third is, HFPN should be extended to handle more advanced type, object that consists of variables and methods. Many parts in a cell, e.g. DNA, mRNA, protein, have known functions, e.g. translation, transcription, degradation, and modification. Thus, if an entity takes the type object that has the methods with these known functions, each process that connects to the entity only needs to call a method of the object in the entity. To realize these three extensions, we have defined two special components; entity and process, called *generic entity* and generic process, respectively. We name this extension of Petri net hybrid functional Petri net with extension (HFPNe) that is defined in Section 2. By extensively using the new features of HFPNe, we model two biological processes that are not normally treated in biopathway modelings, alternative splicing and frameshifting, in Sections 3.1 and 3.2 and two biopathways that contain complicated processes, the pathway of Huntington's disease and the pathway of multi domain modifiable protein p53, in Sections 3.3 and 3.4. All these models with HFPNe demonstrate the new features of HFPNe are important for easy modeling and simulation. In Section 4 discusses the related works of HFPNe by comparing other object based Petri nets, OCPN [19] and Reference nets [18] and our ongoing works.

2 HFPNe: Hybrid Functional Petri Net with Extension

For modeling complex biological processes intuitively, we are required to deal with various kinds of biological information, *e.g.* the density of molecules, the number of molecules, sequences, molecular modifications, binding location, localization of molecules, etc. To cope with this feature in biological system modeling, we introduce *types* for biological entities and processes.

The set T of *types* is defined by the following abstract syntax:

 $\langle type
angle ::=$ boolean || integer || integer+ || real || real+ || string ||

 $pair(\langle type \rangle, \langle type \rangle) \parallel list \langle type \rangle \parallel object(\langle type \rangle, \cdots, \langle type \rangle).$

Then, for $\theta \in T$, we define the *domain* $D(\theta)$ of θ as follows:

- 1. $D(\text{boolean}) = \{\text{true}, \text{false}\}, D(\text{integer}) = \mathbf{Z} \text{ (the set of integers)}, D(\text{integer+}) = \mathbf{N} \text{ (the set of nonnegative integers)}, D(\text{real}) = \mathbf{R} \text{ (the set of real numbers)}, D(\text{real+}) = \mathbf{R}^{\geq 0} \text{ (the set of nonnegative real numbers)}, D(\text{string}) = \mathbf{S} \text{ (the set of strings over some alphabet)}.$
- 2. $D(pair(\theta_1, \theta_2)) = D(\theta_1) \times D(\theta_2).$
- 3. $D(\texttt{list}\theta) = \bigcup_{k>0} D(\theta)^k$.

Table 1: (a) For a connector $c = (e, p) \in EP$, the entity type h(e), the process type h(p) and the connector type a(c)must satisfy the following conditions, where $\sqrt{}$ means that the connection is allowed and - means that the connection is not allowed. (b) For a connector $c = (p, e) \in PE$, the connector type a(c) is **process** by definition. The entity

type h(e) and the process type h(p) must satisfy the following conditions, where $\sqrt{}$ means that the connection is allowed and - means that the connection is not al-

lowed.

connector type		process connector		
pro	cess type	discrete	continuous	generic
entity	discrete	\checkmark	—	\checkmark
type	continuous		\checkmark	\checkmark
	generic	_	—	\checkmark
connector type		associate or		
		inhi	bitory connec	etor
pro	cess type	inhi discrete	bitory connec continuous	tor generic
pro- entity	cess type discrete	inhi discrete $$	bitory connect continuous $$	$\frac{\text{generic}}{}$
pro- entity type	cess type discrete continuous	inhi discrete $$	bitory connect continuous 	$\frac{\text{generic}}{}$
pro- entity type	cess type discrete continuous generic	$\frac{\text{inhi}}{}$	bitory connect continuous 	$\frac{\text{generic}}{}$

connector type		process connector		
pro	cess type	discrete	continuous	generic
entity	discrete	\checkmark	_	\checkmark
type	continuous	\checkmark		\checkmark
	generic		\checkmark	\checkmark
		(b)	1	

4. $D(\texttt{object}(\theta_1, \cdots, \theta_n)) = D(\theta_1) \times \cdots \times D(\theta_n).$ For convenience, we denote $D^* = \bigcup_{\theta \in T} D(\theta).$

Let *E* be a finite set. A type function for *E* is a mapping $\tau : E \to T$. For $e \in E$, $\tau(e)$ is called the type of *e*. A marking of *E* is a mapping $M : E \to D^*$ satisfying $M(e) \in D(\tau(e))$ for $e \in E$. For $e \in E$, M(e) called the mark of *e*. We denote by \mathcal{M} the set of all markings of *E*. We can regard \mathcal{M} as the set $\prod_{e \in E} D(\tau(e))$. Consider a function $f : \mathcal{M} \to \mathbf{R}$. For a subset $F \subseteq E$ and an element $v \in \prod_{e \in F} D(\tau(e))$, let $f[F = v] : \prod_{e \in E - F} D(\tau(e)) \to \mathbf{R}$ be the function obtained from *f* by restricting the value for *F* to *v*, i.e. f[F = v](z) = f(z, v) for $z \in \prod_{e \in E - F} D(\tau(e))$. Let *F* be a subset of *E* such that $e \in F$ satisfies $D(\tau(e)) = \mathbf{R}$ or $\mathbf{R}^{\geq 0}$. We say that the function *f* is continuous for *F* if $f[E - F = v] : \prod_{e \in F} D(\tau(e)) \to \mathbf{R}$ is continuous on $\prod_{e \in F} D(\tau(e))$ for any $v \in \prod_{e \in E - F} D(\tau(e))$.

Based on the above terminology, we define the notion of hybrid functional Petri net with extension (HFPNe). The basic idea of HFPNe is two-fold. The first is to introduce types with which we can deal with various data types. The second is to employ functions of marking f(M) to determine the weight, delay, and speed, etc. which control the system behavior. In the following definition, we use different names instead of place, transition, arc, etc. which are conventionally used in Petri net theory since biological system modeling requires more intuitive names for representing biological entities and processes.

Definition 1 We define a hybrid functional Petri net with extension (HFPNe) $H = (E, P, h, \tau, C, d, \alpha)$ as follows:

- 1. $E = \{e_1, \dots, e_n\}$ is a non-empty finite set of *entities* and $P = \{p_1, \dots, p_m\}$ is a non-empty finite set of *processes*, where we assume $E \cap P = \emptyset^*$.
- 2. $h : E \cup P \rightarrow \{ \text{discrete, continuous, generic} \}$ is a mapping called the *hybrid function* [†] A process $p \in P$ with h(p) = discrete (resp., continuous, generic) is called a *discrete process* (resp., *continuous process, generic process*). An entity $e \in E$ with h(e) = discrete (resp., continuous, generic) is called a *discrete entity* (resp., *continuous entity, generic entity*).

^{*}Terms "entity" and "process" correspond to *place* and *transition*, respectively.

[†]Terms "discrete" and "continuous" correspond to those in hybrid Petri net [1] and "generic" is a newly introduced name which can be of any type in T.

- 3. $\tau : E \to T$ is a type function for E such that $\tau(e) = \text{integer+}$ if e is a discrete entity, and $\tau(e) = \text{real+}$ if e is a continuous entity.
- 4. C = (EP, PE, a, w, u) consists of subsets EP ⊆ E × P and PE ⊆ P × E. An element in EP ∪ PE is called a connector[‡]. Each connector has a connector type which is given by a mapping a : EP ∪ PE → {process, associate, inhibitor} called the connector type function which satisfies the conditions: (i) a(c) = process for c ∈ PE. (ii) All connectors c = (e, p) ∈ EP satisfy the conditions in Table 1(a) and all connectors c = (p, e) ∈ PE satisfy the conditions in Table 1(a) and all connectors c = (p, e) ∈ PE satisfy the conditions in Table 1(a) and all connectors c = (p, e) ∈ PE satisfy the conditions in Table 1(a) and all connectors c = (p, e) ∈ PE satisfy the conditions in Table 1(b). A connector c = (e, p) ∈ EP is called a process connector (resp., an associate connector, an inhibitory connector) if a(c) = process (resp., associate, inhibitor)[§]. For a connector c = (p, e) ∈ PE, a(c) = process by definition and we also call it a process connector. We say that a connector c = (e, p) ∈ EP is discrete (resp., continuous, generic) if p is a discrete process (resp., continuous process, generic process). Let M be the set of all markings of E and let F be the set of continuous entities in E. Then we denote D_{discrete} = {f | f : M → N}, D_{continuous} = {f | f : M → R^{≥0} is continuous for F}, D_{generic} = {f | f : M → D^{*}}, and D_{boolean} = {f | f : M → {true, false}}. Then w and u are given as follows:
 - (a) $w: EP \to \mathcal{D}_{\text{discrete}} \cup \mathcal{D}_{\text{continuous}} \cup \mathcal{D}_{\text{boolean}}$ is a function called the *activity function* such that for a connector $c \in EP$ (i) $w(c) \in \mathcal{D}_{\text{discrete}}$ if c is discrete, (ii) $w(c) \in \mathcal{D}_{\text{continuous}}$ if c is continuous, (iii) $w(c) \in \mathcal{D}_{\text{boolean}}$ if c is generic. For a connector (e, p), w(e, p) be used as a function giving the threshold in discrete and continuous cases and the condition in generic case which is required for enabling the process p.
 - (b) $u: EP \cup PE \to \mathcal{D}_{\text{discrete}} \cup \mathcal{D}_{\text{continuous}} \cup \mathcal{D}_{\text{generic}}$ is a function called the *update function* which satisfies the following conditions: For a connector $c \in EP \cup PE$, let $c = (e, p) \in EP$ or $c = (p, e) \in PE$. (i) $u(c) \in \mathcal{D}_{\text{discrete}}$ if c is discrete. (ii) $u(c) \in \mathcal{D}_{\text{continuous}}$ if c is continuous. (iii) If c is generic, then u(c) is a function in $\mathcal{D}_{\text{generic}}$ such that u(c)(M) is in $D(\tau(e))$ for any marking $M \in \mathcal{M}$. For a connector c = (e, p) or c = (p, e), u(c) is used as a function which will update the mark of e.
- 5. $d: P_{\text{discrete}} \to \mathcal{D}_{\text{continuous}}$ is a mapping called the *delay*, where P_{discrete} is the set of discrete processes in P. For a discrete process $p, d(p) : \mathcal{M} \to \mathbf{R}^{\geq 0}$ is called the *delay function* of p.
- 6. $\alpha > 0$ is a real number called the *generic time*. The generic time is used as the clock for generic processes.

For graphical representation, HFPNe inherits the tradition of other Petri nets as in Figure 1.

We introduce a parameter $t \in \mathbf{R}^{\geq 0}$ called the *time* to a hybrid functional Petri net with extension $H = (E, P, h, \tau, C, d, \alpha)$. Given a marking I called the *initial marking*, we define a marking M(t) called the *marking at time* t and a marking $M_r(t)$ called the *reserved marking at time* t for $t \geq 0$ in the following way. By convention, we denote M(e,t) = M(t)(e) and $M_r(e,t) = M_r(t)(e)$ for $e \in E$. We define $\tilde{M}(t)$ by $\tilde{M}(e,t) = M(e,t) - M_r(e,t)$ for discrete and continuous entities and $\tilde{M}(e,t) = M(e,t)$ for generic entities e.

First, we define M(0) = I, $M_r(e, 0) = 0$ for all discrete and continuous entities e. For all generic entities e, $M_r(e, t) = \text{null}$ (the empty list) for any $t \ge 0$. For t > 0, we define M(t) and $M_r(t)$ in the following way. For a process $p \in P$ at time t, if the following conditions are satisfied, then the process p is said to be *enabled* at time t. Otherwise the process is said to be *unenabled* at time t.

[‡] "Connector" corresponds to *arc*.

[§] "Process connector", "associate connector" and "inhibitory connector" correspond to *arc with weight, test arc* and *inhibitory arc*, respectively.

- 1. If p is a discrete process, then for all connectors $c = (e, p) \in EP$ the following conditions hold:
 - (a) $\tilde{M}(e,t) \ge w(e,p)(M(t))$ if $a(c) \ne \text{inhibitor}$.
 - (b) $\tilde{M}(e,t) < w(e,p)(M(t))$ if a(c) = inhibitor.
- 2. If p is a continuous process, then for all connectors $c = (e, p) \in EP$ the following conditions hold:
 - (a) $\tilde{M}(e,t) \ge w(e,p)(M(t))$ if $a(c) \ne inhibitor$.
 - (b) $\tilde{M}(e,t) \leq w(e,p)(M(t))$ if a(c) = inhibitor.
- 3. If p is a generic process, then for all connectors $c = (e, p) \in EP$ the following conditions hold:
 - (a) $w(e,p)(\tilde{M}(t)) =$ true if $a(c) \neq$ inhibitor.
 - (b) $w(e,p)(\tilde{M}(t)) =$ false if a(c) =inhibitor.

If an unenabled process turns to be enabled at time t, the process is said to be *triggered* at time t. If an enabled process turns to be unenabled or an unenabled process turns to be enabled at time t, the process is said to be *switched* at time t. If a discrete process p is triggered at time t, we say that the discrete process can be *fired* at time t + d(p)(M(t)). If a generic process p is triggered at time t, we say that the generic process can be *fired* at time $t + \alpha$.

For an entity $e \in E$ and time t, let $S_d(t)$ be the set of discrete processes which can be fired at time t, and let $U_d(t)$ be the set of discrete processes which are triggered at time t. For a discrete process p that can be fired at time t, we denote by q(p,t) the time when p is triggered. Let $S_c(t)$ be the set of continuous processes which are enabled at time t. Let $S_g(t)$ be the set of generic processes which can be fired at time t.

Note that we can choose a sufficiently small $\varepsilon_t > 0$ such that in the interval $[t - \varepsilon_t, t)$, neither discrete nor generic process is triggered or can be fired and no continuous process is switched.

Also note the following facts:

- 1. $S_c(t \varepsilon_t) = S_c(t')$ for any $t' \in [t \varepsilon_t, t)$ since no continuous process is switched in the interval $[t \varepsilon_t, t)$.
- 2. $\hat{M}(t')$ is constant on $E E_{\text{continuous}}$ in the interval $[t \varepsilon_t, t)$ since neither discrete nor generic process is triggered or can be fired in the interval $[t \varepsilon_t, t)$, where $E_{\text{continuous}} = \{e \in E \mid e \text{ is continuous}\}$.
- 3. For any continuous connector c, $u(c)(\tilde{M}(t'))$ is continuous on $[t \varepsilon_t, t)$ since by definition u(c) is continuous for $E_{\text{continuous}}$ and $\tilde{M}(t')$ is constant on $E E_{\text{continuous}}$ in the interval $[t \varepsilon_t, t)$.

Then M(t) is defined by the following procedure:

- 1. $Tmp \leftarrow M(t \varepsilon_t), Tmp_r \leftarrow M_r(t \varepsilon_t)$ 2. **if** $t = \alpha k$ for some integer $k \ge 1$ **then for each** generic process $p \in S_g(t)$ $Tmp' \leftarrow Tmp$ **for each** $(e, p) \in EP$ with a(e, p) = process $Tmp'(e) \leftarrow u(e, p)(Tmp)$ **for each** $(p, e) \in PE$ $Tmp'(e) \leftarrow u(p, e)(Tmp)$ $Tmp \leftarrow Tmp'$
- 3. for each continuous process $p \in S_c(t \varepsilon_t)$

 $Tmp' \leftarrow Tmp$ for each $(e, p) \in EP$ with a(e, p) = process $Tmp'(e) \leftarrow Tmp'(e) - \int_{t-\varepsilon_t}^t u(e, p)(\tilde{M}(x))dx$ for each $(p, e) \in PE$ $Tmp'(e) \leftarrow Tmp'(e) + \int_{t-\varepsilon_t}^t u(p, e)(\tilde{M}(x))dx$ $Tmp \leftarrow Tmp'$ 4. for each discrete process $p \in S_d(t)$ $Tmp' \leftarrow Tmp$ for each $(e, p) \in EP$ with a(e, p) = process $Tmp'(e) \leftarrow Tmp'(e) - u(e, p)(\tilde{M}(q(p, t)))$ for each $(p, e) \in PE$ $Tmp'(e) \leftarrow Tmp'(e) + u(p, e)(\tilde{M}(q(p, t)))$ $Tmp \leftarrow Tmp'$ 5. $M(t) \leftarrow Tmp$

Then $M_r(t)$ is defined as follows:

6. for each entity
$$e$$
 with $h(e) = \text{discrete or continuous}$
 $Tmp_r(e) \leftarrow Tmp_r(e) - \sum_{\substack{p \text{ with } p \in S_d(t) \\ \text{and } (e,p) \in EP}} u(e,p)(\tilde{M}(q(p,t))) + \sum_{\substack{p \text{ with } p \in U_d(t) \\ \text{and } (e,p) \in EP}} u(e,p)(\tilde{M}(t-\varepsilon_t)).$

7. $M_r(t) \leftarrow Tmp_r$.

We call M(t) $(t \ge 0)$ the behavior of H starting at the initial marking M(0) = I.

2.1 Remarks in Implementation of HFPNe

In Genomic Object Net, we have implemented a simulator of HFPNe by approximating the time t by $t = 0, \delta, \dots, k\delta, \dots$ for integers k by using an appropriately small real number $\delta > 0$. Furthermore, the generic time α is also set to be $\delta = \alpha$ for simplicity.

Another issue in implementation is the problem of conflict resolution. In the above procedure, Step 2 for generic processes and Step 4 for discrete processes may have conflicts for execution. Let p_{i_1}, \ldots, p_{i_t} be processes which can be fired in Step 2 or Step 4. In our implementation, we arrange these processes in a random order and execute the processes according to this order. During this execution, we will skip the processes which cannot be fired anymore due to the changes of marks of entities.

2.2 Relationships with Other Petri Nets

HFPN $H = (E, P, h, \tau, C, d)$ is defined from HFPNe by deleting all matters with "generic" and by adding the restriction that u(c) = w(c) for any discrete connector $c = (e, p) \in EP$. This condition means that the weight w(c) of the connector is the same as the number of tokens u(c) removed from the entity e by firing. This convention is traditionally employed in Petri net as a weight. In our definition of HFPNe, however, we have separated these two notions.

HPN $H = (E, P, h, \tau, C = (EP, PE, a, w, u), d)$ [1] is defined by adding the following restriction to HFPN: (i) w(c), u(c), and d(p) are constants for any connector $c \in EP \cup PE$ and any process $p \in P$. HDN $H = (E, P, h, \tau, C = (EP, PE, a, w, u), d)$ [10, 11] is also defined by adding the following

DNA stepl transcription precurson P.P. EX mRNA step2 5'capping 7-metylqu step3 alternative splicing m'Gp-p-p Exon1 E m'GPPPP EXON2 EXON3 3'polyadenilation step4 m'GPPP EXONI EXON2 E A A --- A 250 m'GPPP EXON2 EXON3 RP -(A) (A) (A) - - (A) 251 Calcitionin CGRP

Figure 2: Alternative RNA splicing model of the *Calcitonin/CGRP* gene in Figure 3.



Figure 3: An alternative RNA splicing model of the *Calcitonin/CGRP* gene with HFPNe.

restriction to HFPN: (ii) For any continuous process p, it is assumed that w(e, p) = u(p, e') for any process connectors $(e, p) \in EP$ and $(p, e') \in PE$.

If we delete all matters with "discrete" from HPN, we have the definition of CTPN. If we delete all matters with "continuous" from HPN, we have the definition of TPN. Furthermore, if we delete the matters with "delay" from TPN, we have the definition of the original Petri net. Thus, HFPNe is a highly abstract extension of Petri net for biological process modeling which can involve PN, TPN, CTPN, HPN and HFPN as its special cases.

3 Modeling of Biological Processes with HFPNe



Figure 1: Graphical notations of HFPNe components with conventional way.

HFPNe models as in Figures 3, 4(a), 5 and 6.

In this section, we demonstrate that more biological processes can be easily modeled with HF-PNe than HFPN. For this purpose, we selected four biological processes for modeling that extensively use the HFPNe features. These biological processes are important activities in living cells and should be handled with application tools that aim to model and simulate biological systems. Our aim is not only to theoretically describe how to model biological processes but also to provide a useful application tool to users in biology and medicine. We have developed GON [9, 27] which is based on the HFPNe architecture. In the following sections, we exemplify with GON that complex biological processes can be modeled while illustrating snapshots of

3.1 From DNA to mRNA in Eucaryotes - Alternative Splicing

The mechanism from DNA to mRNA in eucaryote is more complicated than that in bacteria. The major difference derives from eucaryotic genes which consist of two regions, i.e. exons and introns. As in Figure 2, the mechanism mainly consists of four steps; **step1**: transcription, **step2**: 5' capping, **step3**: RNA splicing, and **step4**: 3' polyadenylation. In **step1**, DNA is transcribed into precursor-mRNA (specially named to distinguish from mRNA (mature-mRNA) after **step4**). In **step2**, 5' end of

conn-	connector	r update fu	update function		
ector	type				
a01	process	import("go	<pre>import("gon.Transcription");</pre>		
		<pre>totalnum = m08.length(); num = m01.length();</pre>		ength();	
		if(totalnu	um > num){		
		nextcode =	m08.substrin	g(totalnum-nur	n-1,totalnum-num);
		newsequend	ce = m01 + Tra	nscription::T	rans(nextcode);}
		else{new_s	sequence = "";	} return newse	equence;
a02	process	import("go	on.Transcripti	on");	
		if(Transci	ription::Finis	h(m01,m08)){re	eturn m01;} else{return "";}
a03	process	return m02	2;		
a04	process	import("go	on.Transcripti	on");	
		if(!m02.ed	<pre>quals("")){ret</pre>	urn Transcript	<pre>tion::Capping(m02);} else{return "";}</pre>
a05	process	return "";			
a06	process	import("go	on.Splicing_Ca	lcitoninCGRP")); if(m03.equals("")){return m03;}
		<pre>else{return Splicing_CalcitoninCGRP::AlternativeSplicing(m03);}</pre>		:AlternativeSplicing(m03);}	
a08	process	return "";	return "";		
a10	process	return m10	return m10;		
a11	process	<pre>num = 0; for(i=0;i<m05.length;++i){if(m05[i].equals(m09)){num++;}}< pre=""></m05.length;++i){if(m05[i].equals(m09)){num++;}}<></pre>		f(m05[i].equals(m09)){num++;}}	
		return num	1;		
a12	process	return m05	;		
a13	process	num = 0; f	for(i=0;i <m05.< td=""><td><pre>length;++i){if</pre></td><td>f(m05[i].equals(m10)){num++;}}</td></m05.<>	<pre>length;++i){if</pre>	f(m05[i].equals(m10)){num++;}}
		return num	1;		
a14	process	return m05	return m05;		
a15	process	<pre>import("gon.Transcription"); if(m04.equals("")){return m05;}</pre>		equals("")){return m05;}	
<pre>else{return m05+[Transcription::Polyadenylation(m04)];}</pre>		adenylation(m04)];}			
entity		entity type	type	initial mark	Table 3: Properties of enti-
m01/m0	2/m03/m04	generic	string	(())	tion for the Calaitanin/CCPP
m05		generic	list string	()	
m06/m0	7	discrete	integer+	0	transcription model in Fig-
m08/m09/m10		generic	string	see [43]	ure 3.

Table 2: Update functions of connectors for the *Calcitonin/CGRP* transcription model in Figure 3. External Java classes Transcription, Splicing and Splicing_CalcitoninCGRP are in our site [43].

the precursor-mRNA is modified. In **step3**, each splicing event removes one intron and accordingly all introns of the precursor-mRNA are removed. Finally in **step4**, 3' end of precursor-mRNA is modified to produce mature-mRNA in order to allow the cell to assess whether both ends of the mRNA are present before it exports the RNA sequence from the nucleus for translation into proteins.

In **step3**, there is a eucaryote specific splicing process named alternative RNA splicing. The alternative RNA splicing is to produce different mRNA from the same precursor-mRNA by splicing it in different ways. The DNA to mRNA transcription with alternative RNA splicing was found in 1982 for *Calcitonin/CGRP* gene expression [2]. We draw focus to model *Calcitonin/CGRP* gene expression while combining other recent biological knowledge [7, 36].

As in Figure 2, the *Calcitonin* gene expression consists of four introns and five exons, and the transcription process progresses with **step1**, **step2**, **step3** and **step4**. In **step3**, by alternative splicing events, one of exon1/exon2/exon3/exon4 (say set1) or exon2/exon3/exon5/exon6 (say set2) is selected. If set1 (set2) is selected, the mature-mRNA is translated into *Calcitonin* (*CGRP*), respectively. The HFPNe model can faithfully realize these steps as generic processes and realize precursor-mRNA and mature-mRNA states as generic entities with the type **string** (see Figure 3). In the model, the mature-RNAs, *Calcitonin* mRNA and *CGRP* mRNA, are represented by the generic entity m09 with the type **list string**. The generic entities, m12,m13,m14, and m15 are used to inform which sequence should be spliced in the generic processes, t03,t04,t05, and t06, respectively. These generic notions are necessary for modeling these four steps from DNA to mRNA. It is hard to model with HFPN [9]

and other simulation tools [26, 35]. The detailed parameters and functions are described in Tables 2 and 3.

As another example of alternative splicing, *DSCAM* gene in *Drosophila*, where for exons (named exon A, B, C, and D) are selected and combined. Each precursor-mRNA contains 12 alternatives for exon A, 48 alternatives for exon B, 33 alternatives for exon C, 2 alternatives for exon D [38]. Thus, there are 38,016 possible mature mRNAs for the DSCAM gene. If the model is created with HFPN, we have to deal with 38,016 entities for this DSCAM transcription process. If the model is created with HFPN, 38016 entities are necessary for the simple *DSCAM* transcription model.

3.2 Translation of mRNA - Frameshift

Frameshift is also an important biological process that occurs during RNA translations. The frameshift is to skip or reread some ribonucleotides when translating RNAs and it is commonly utilized by many RNA viruses as programmed ribosomal frameshifting [14].

For example, a model of ribosomal frameshifting in the human immunodeficiency virus (HIV-1) (gag-pol expression in Figure 4(b)) can be modeled with HFPNe as in Figure 4(a). As in Figure 4, two proteins, gag and pol, are produced from one RNA sequence with the frameshift by rereading a ribonucleotide, adenine (A), twice. In HFPNe, the generic entity e3 has the type pair(integer (i1), string (s1)), in order to express frameshifting states; i1 denotes how many ribonucleotides are skipped (+1) or reread (-1) and s1 denotes the length of current translated amino-acid sequence. The frameshifting occurs at a specific point. In addition the probability of reread at the point is 0.1 in wild type [14]. Thus, in the model we assign an mRNA translation skip function with stochastic behavior to this specific point. The stochastic feature is already realized in HFPN and HFPNe also inherits this feature. The detailed update functions are summarized in Table 4(c) and the initial marks and types of entities are summarized in Figure 4(d). It is also not straightforward to represent the ribosomal frameshifting with HFPN.

3.3 Huntington's Disease

As an advanced pathway model with HFPNe, we selected a genetic disease called Huntington's disease. Because in biology and medical sciences disease pathway modeling and simulation are becoming one of the most important topics to treat the disease from the pathway level. Huntington's disease is an autosomal dominant progressive neurodegenerative disorder that is characterized by chorea, psychiatric disturbances, and dementia [20]. The disease is in short the generic defects of *Huntingtin*. *Huntingtin* is a multi-domain protein with a polymorphic glutamine/proline (G/P)-rich domain at the N terminus [37]. Polyglutamine (polyQ) sequences in unaffected individuals range from 11 to 34 glutamine residues, whereas those of *Huntingtin* disease patients contain 37 or more glutamine residues (more than 90)[4]. The disease appears when a specific polyQ length is exceeded.

Based on the disease model proposed by Wellington *et al.* [13], we created a HFPNe model for this disease in Figure 5. Our model uses following known experimental facts in the literature.

- fact1 Huntingtin can be cleaved by caspase-3 and yields two fragments, N terminal region (NT) that contains polyQ repeats in Huntingtin and C terminal region (CT) [13].
- fact2 Both of *Huntingtin* disease and normal *Huntingtin* can cleave and the rates of fragmentation are the same [40].
- fact3 *Procaspase-3* has low-level catalytic activity for disease *Huntingtin* and is capable of cleaving the same substrates, i.e. *NT* and *CT*, as activated *caspase-3* [32].
- fact4 When cleaved by caspase-3, NT has a responsibility for cytotoxicity [15, 41].
- fact5 NT has the ability to form protein aggregates and can be found in the nucleus and in the cytoplasm [33].



(;	a)
(IJ

connector	update function
a3	<pre>m4 = 0;for(i=0;i<m3.length;++i){if(m3[i].equals(m6)){m4++;}}return m4;<="" pre=""></m3.length;++i){if(m3[i].equals(m6)){m4++;}}return></pre>
a4	<pre>m5 = 0;for(i=0;i<m3.length;++i){if(m3[i].equals(m7)){m5++;}}return m5;<="" pre=""></m3.length;++i){if(m3[i].equals(m7)){m5++;}}return></pre>
a5	<pre>import("gon.Translation");</pre>
	num=m2[1].length(); n=m1.substring(num*3+m2[0],(num+1)*3+m2[0]);
	<pre>if(Translation::EndCode(n)){ m3 += [m2[1]]; }return m3;</pre>
a7	<pre>import("gon.Translation");</pre>
	num=m2[1].length(); n=m1.substring(num*3+m2[0],(num+1)*3+m2[0]);
	<pre>if(!Translation::EndCode(n)){m2[1] += Translation::Trans(n);</pre>
	if(num==2 && Math::random()<=0.1){m2[0]=-1;}}
	else{m2[0] = 0;m2[1] = "";}return m2;

1		<u>۱</u>
1	\mathbf{r}	۱.
١.	t,	
1		/

entity	entity type	type	initial mark
m1/m6/m7	generic	string	see $[43]$
m2	generic	<pre>pair(integer,string)</pre>	(0, "")
m3	generic	list string	()
m4/m5	discrete	integer+	0

(d)

Figure 4: (a) Ribosomal frameshifting in HIV-1 gag-pol. (b) Ribosomal frameshifting in HIV-1 gagpol expressed with HFPNe. The characterization of ribosomal frameshifting was first described by Tyler [14]. (c) Update functions in (b). (d) Properties of entities in (b).

fact6 The aggregate number of NT increases in proportion to the length of polyQ [34].

fact7 NT fragments of mutant Huntingtin have the ability to induce caspase-3 [6].

fact8 Both of disease and normal NT can cross the nuclear membrane. The smaller the cleavage product, the greater the tendency of *Huntingtin* cross [12, 39].

Huntingtin and NT change their function depending on their polyQ length (fact3, fact6, fact7). Thus, in our model, these proteins are represented with generic entities with the type: pair(integer+, real+). The first attribute integer+ corresponds to the length of polyQ and the second attribute real+ corresponds to the quantity of the protein, i.e. NT or Huntingtin. In our model, NT and Huntingtin are represented with m1 and m4, respectively. The properties of processes and entities in the model are summarized in Tables 4 and 5. Huntingtin m4 is translated by the generic process t09 while procaspase-3 m6 is translated by the continuous process t10. The procaspase-3 m6 can cleave t12 disease Huntingtin (fact3). Thus, the weight function of associate connector a14 has a function that depends on the length of huntingtin m4 and cleavage process t12 is not enabled for normal Huntingtin.

(b)

process	process type
t01/t02/t04/t06/t07/t08/t10/t17/t18	continuous
t03/t05/t09/t11/t12/t13/t14/t15/t16/t19	generic

entity	entity	type	initial
	type		mark
dt	continuous	real+	0.0
m1/m2/m4	generic	<pre>pair(integer+</pre>	(n, 0.0)
		,real+)	
m3/m5/m6/m7/m8/m9	continuous	real+	0.0

Table 4: Properties of processes for Huntington's disease model in Figure 5.

Table 5: Properties of entities for the huntingtin disease model in Figure 5. Initial value n in m1, m2, and m4 denotes the length of Q repeats in the huntingtin protein and must be assigned an integer value when simulating the model.

The activity functions and types of connectors are summarized in Table 6. On the other hand, the *caspase-3* m5 can cleave normal *Huntingtin* and disease *Huntingtin* that is modeled with the process t11 (fact2). Thus, the weight of associate connector a21 has a function that does not depend on the length of *Huntingtin* m4. *Huntingtin* m4 is cleaved and separated into CT m3 and NT m1 (fact1). CT does not contain the polyQ region and is represented with continuous entity m3 (fact1).

NT inherits the polyQ region. The length of NT relates to the activity of the process t13 that changes *procaspase-3* m6 into *caspase-3* m5 (fact7). To model it, the weight function of associate connector a15 depends on the length of NT m1 and process t13 can not be activated by normal huntingtin.

NT in cytoplasm migrates to nucleus (fact8), and NT in nucleus m2 causes cell death t19 with the cytotoxicity (fact4). From fact5, cell death can be prevented by aggregation of NT. Moreover, the aggregation rate depends on the length of polyQ (fact6). NT in nucleus m2 should therefore be denoted by generic entities such as *Huntingtin* m4 and NT m1 in cytoplasm. NT in the cytoplasm m1 and NT in the nucleus m2 connect to the aggregation process t15 and t16, respectively. The update functions of process connectors a23 and a26 represent aggregate speeds and depend on the length of polyQ in NT in nucleus m2 (fact6). The update functions of connectors are summarized in Table 7.

With the model, we can simulate the *n* polyQ length huntingtin $(0 \le n \le 100 [4])$ by only changing the initial marks of m1, m2, and m4 as in Table 5. If the same disease model is created with HFPN, at least extra (i) one entity and (ii) many connectors are necessary; (i) an entity (say e1) that denotes the length of huntingtin, (ii) associate connectors that point from e1 to every process that has more than one connector whose weight function or speed function depends on the value of e1. Thus, the HFPNe huntingtin disease model is simpler than the HFPN model.

3.4 Protein Modification - p53

The previous HFPNe models extensively use generic entities and generic processes. However, their types of generic entities are only simple ones, *e.g.* string, list real, list string, pair(integer+, string+), pair(integer+, real+). A generic entity can take more advanced type object as its type. Under HFPNe application GON, an object corresponds to a Java *class* and a *method* of an object is a specific function assigned to the object. If a generic entity has the type object, the entity can be initialized with an object with methods and variables and these methods can be applied to update functions of connectors that join the generic entity and generic processes, and change the mark by updating variables of the entity.

To show the effectiveness of generic entities with the type object, we deal with protein modification biological processes - *e.g.* phosphorylation, acetylation, and methylation - with enzymes. Specifically, the protein p53 that can be modified by enzymes - CK1, ATM, DNAPK, TFIIH, MAPK, PCAF, PKC,



Figure 5: Huntington's disease model with HFPNe. HFPNe components in this figure are replaced with suitable images that represents biological information.

p300, CK2, and JNK - is modeled [16]. By these enzymes, eighteen positions of p53 are modified. There are 2^{18} patterns. Thus, it is again hard to create a model with HFPN because 2^{18} entities are necessary for the model with HFPN.

To model with HFPNe while using the type object for generic entities, Protein and Enzyme classes are created and these classes are inherited in order to realize specific Protein class, *e.g.* p53, and Enzyme classes, *e.g.* CK1, ATM, DNAPK, TFIIH, MAPK, PCAF, PKC, p300, CK2, and JNK. As in Table 9, (i) the generic entity m01 takes a specific object ProteinSet that is initialized with p53 object that corresponds to p53 class and inherits Protein class, (ii) the generic entities m02, m03, m04, m05, m06, m07, m08, m09, m10, and m11 take specific objects whose classes inherit Enzyme classes CK1, ATM, DNAPK, TFIIH, MAPK, PCAF, PKC, p300, CK2, and JNK, respectively (a class name corresponds to an enzyme name). These objects are implemented with Java classes on GON. Full codes of these classes are available from [43].

To describe modification biological processes by enzymes, the generic processes t01,..., t10 are used. As in Table 10, the update functions a13,a14,a15,a16,a17, a18,a19,a20,a21,a22 of the generic processes call the Modify method of p53 class to simulate the modification biological process by CK1, ATM, DNAPK, TFIIH, MAPK, PCAF, PKC, p300, CK2, JNK, respectively. To describe translation and degradation biological processes of p53, the generic processes t11 and t12 are created, respectively. As in Table 10, processes t11 and t12 follow connectors with update functions a21 and a22 that call Translation and Degradation methods of p53 class to simulate the translation and degradation biological processes, respectively.

At the first glance, HFPNe model of p53 modification biological processes seems complicated. However, as in Table 10, in the p53 model, all update functions just call a specific method, *e.g.* Modify,



Figure 6: A protein modification model of the p53 protein with HF-The generic en-PNe. tity in the center m01 is modification target p53and other generic entities $m02, \cdots, m11$ are enzymes that modify the p53 en-Generic processes tity. $t01, \cdots, t10$ have connectors $a01, \dots, a10$ and by their update functions, the modified state of p53in m01 is updated.

Translation, and Degradation. As in Table 9, all initial marks of generic entities are defined by just calling the specific method, i.e. Initialize. Thus, if objects with suitable methods that correspond to biological processes are created as a biopathway library, the majority of users, biological and medical scientists without programming skills, can create biological models. But with their expert knowledge of biological processes, they can create models by simply assigning a suitable object to a generic entity and a suitable method to the update function of the generic connector.

We can apply the same approach to the previous biological processes. In the previous example, generic entities can take type object whose class is DNA with transcription method or RNA with translation method instead of string primitive type. Thus, owing to the object extension of HFPNe, more complicated biological processes can be easily modeled than with HFPN.

On the other hand, the Gene Ontology (GO) Consortium is trying to produce a controlled vocabulary that can be applied to all organisms as knowledge of gene and protein roles in cells [3]. By applying these gene ontology related vocabularies to names of objects and their methods, we can create a more standardized library for biological processes with HFPNe.

connector	connector type	activity function
a02/a09/a19/a20	process	0.0
a04/a05/a06/a11	process	return true;
a12	process	return {(m4[1]>20.0) ? true : false;}
a14	associate	return {(m4[0]>37&&m6>0.0) ? true : false;}
a15	associate	return {(m4[0]>37&&m1>0.0) ? true : false;}
a17	process	20.0
a21	associate	return {(m5[1]>0.0) ? true : false;}
a23/a26/a29	process	return true;
a25/a28	process	1.0
a31	process	return {(m2[1]>10.0) ? true : false;}

Table 6: Properties of connectors for Huntington's disease model in Figure 5. A property of connectors, update function is summarized in Table 7.

connector	connector type	update function
a01/a02	process	1.0
a03	process	m4[1] = m4[1]+1.0*dt; return m4;
a04	process	m4[1] = m4[1] - (m4[1]*dt)/200; return m4;
a05	process	m4[1] = m4[1]-0.0001*m4[1]*m6*dt; return m4;
a06	process	m4[1] = m4[1]-0.001*m4[1]*m5*dt; return m4;
a07	process	m3 = m3+0.0001*m4[1]*m6*dt; return m3;
a08	process	m3 = m3+0.001*m4[1]*m5*dt; return m3;
a09	process	m3/50
a10	process	m1[1] = m1[1]+0.001*m4[1]*m5*dt; return m1;
a11	process	m1[1] = m1[1]-(m1[1]/100)*dt; return m1;
a12	process	m1[1] = m1[1]-0.001*m1[1]*dt; return m1;
a13	process	m2[1] = m2[1]+0.001*m1[1]*dt; return m2;
a16	process	1.0
a17	process	m6 = m6 - 0.001*m1*m6*dt; return m6;
a18	process	m5 = m5 + 0.001*m1*m6*dt; return m5;
a19	process	m6/100
a20	process	m5/100
a22	process	m1[1]=m1[1]+0.0001*m4[1]*m6*dt;return m1;
a23	process	m1[1]=m1[1]-(m1[1]*(m1[0]/1000))*dt;return m1;
a24	process	m7=m7+(m1[1]*(m1[0]/1000))*dt;return m7;
a25	process	m7/100
a26	process	m2[1]=m2[1]-(m2[1]*(m2[0]/1000))*dt;return m2;
a27	process	m8=m8+(m2[1]*(m2[0]/1000))*dt;return m8;
a28	process	m8/100
a29	process	m2[1]=m2[1]-(m2[1]/100)*dt;return m1;
a30	process	m9+=0.001*m2[1];return m9;

Table 7: Update functions of connectors for the huntingtin disease model in Figure 5. Update function must be defined for all process connectors that are connected with at least one generic process.

Table 8: The protein modification positions and modification type of p53 enzymes [16].

enzyme	modification	modification	
	\mathbf{type}	positions	
CK1	Phosphorylation	S4,S6,S9,S18	
ATM	Phosphorylation	S15	
DNAPK	Phosphorylation	S15,S37	
TFIIH	Phosphorylation	S33,S315	
		S378,S392	
MAPK	Phosphorylation	T73,T83	
PCAF	Acetylation	K320	
PKC	Phosphorylation	S378	
p300	Acetylation	K382	
CK2	Phosphorylation	S392	
JNK	Phosphorylation	S33	



entity	entity type	type	initial mark
m01	generic	object	<pre>ProtenSet::Initialize(m01,p53::Initialize())</pre>
m02	generic	object	CK1::Initialize(m02)
m03	generic	object	ATM::Initialize(m03)
m04	generic	object	DNAPK::Initialize(m04)
m05	generic	object	TFIIH::Initialize(m05)
m06	generic	object	MAPK::Initialize(m06)
m07	generic	object	PCAF::Initialize(m07)
m08	generic	object	PKC::Initialize(m08)
m09	generic	object	p300::Initialize(m09)
m10	generic	object	CK2::Initialize(m10)
m11	generic	object	JNK::Initialize(m11)

Table 9: Properties of entities for the p53 modification model in Figure 6.

conn-	connector	update
ector	type	function
a11	process	<pre>m01.Translate(); return m01;</pre>
a12	process	<pre>m01.Degradation();return m01;</pre>
a13	process	<pre>m01.Modify(m02); return m01;</pre>
a14	process	<pre>m01.Modify(m03); return m01;</pre>
a15	process	<pre>m01.Modify(m04); return m01;</pre>
a16	process	<pre>m01.Modify(m05); return m01;</pre>
a17	process	<pre>m01.Modify(m06); return m01;</pre>
a18	process	<pre>m01.Modify(m07); return m01;</pre>
a19	process	<pre>m01.Modify(m08); return m01;</pre>
a20	process	<pre>m01.Modify(m09); return m01;</pre>
a21	process	<pre>m01.Modify(m10); return m01;</pre>
a22	process	<pre>m01.Modify(m11); return m01;</pre>

Table 10: Update functions of the *p53* modification model with HF-PNe in Figure 6. All update functions are written in Pnuts language [44]. External java classes, CK1, ATM, DNAPK, TFIIH, MAPK, PCAF, PKC, p300, CK2, JNK, ProteinSet, Enzyme, and Protein are described in [43]. Connectors a01,.,a10 have connector type associate and update function do not exist.

4 Discussion and Conclusion

In this paper, new versatile Petri net based architecture HFPNe is demonstrated for easy biological process modeling and simulation. HFPNe introduces the notions of generic entities and generic processes to HFPN. In HFPNe architecture, generic entities can hold various kinds of types including object. With the feature and inherited features of HFPN, complex biological processes can be effectively modeled. Four biological processes, alternative splicing, frameshifting, Huntington's disease, and multi-domain modification process of p53 are employed in order to show the effectiveness of HFPNe.

From the theoretical point of view in the Petri net community, HFPNe can be one of the object oriented Petri nets (OPN). This approach was first given by Becker and Colom [5]. In the community, two advanced Petri nets based on the OPN architectures have been proposed, Objective Colored Petri Nets (OCP-Nets) [19] and Reference nets [18]. OCP-Nets is based on the OPN enhanced by the fusion place concept of Hierarchical Colored Petri Nets (HCPN). With this concept, OCP-Nets can dynamically change (shrink or grow) their structure of the net in simulation, the feature does not exist in HFPNe. Reference nets is also based on the OPN enhanced by the reference entities concept. With the concept, (i) an entity in a net (net1) can hold another net (net2), (ii) the net2 can move to other entities in the net1 by firing, (iii) the net2 can also change its state by firing. The differences between them are as follows: Reference nets can move the net but OCP-Nets cannot move the net in simulation processes. Besides OCP-Nets can create new connection rules in simulation processes. The HFPNe does not support these two OCP-Nets and Reference nets features. On the other hand, the feature of delay function of HFPNe does not exist in these object oriented Petri nets. However, in a cell, various kinds of time scale biological processes exist and to model these biological processes in one model, the notion of delay is necessary. With delay functions, HFPNe can handle these biological processes efficiently. In theoretical point of view, it may be easy to extend HFPNe to support OCP-Nets or Reference nets features because an implementation of HFPNe GON has been already extended to treat the feature to add/remove entities and processes while simulation.

As the reason that is already described in Section 3.4, we are trying to create a biological process library with HFPNe on GON. However, it is difficult to create all biological processes at once. Thus, as the first step, all processes in Kyoto Model [24] will be recompiled as a part of the library. Kyoto Model is a ventricular cell model by compiling classical electro-physiological findings. In the process of reconstruction, we will persist efforts to locate the best approach to systematically reconstruct existing other biological process models like Kyoto Model. Then we will apply the systematic approach to these other biological process models for enriching the library.

References

- Alla, H. and David, R., Continuous and hybrid Petri nets, Journal of Circuits, Systems, and Computers, 8:159–188, 1998.
- [2] Amara, S.G., Jonas, V., Rosenfeld, M.G., Ong, E.S., and Evans, R.M., Alternative RNA processing in calcitonin gene expression generates mRNAs encoding different polypeptide products, *Nature*, 298:240–244, 1982.
- [3] Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T., Harris, M.A., Hill, D.P., Issel-Tarver, L., Kasarskis, A., Lewis, S., Matese, J.C., Richardson, J.E., Ringwald, M., Rubin, G.M., and Sherlock, G., Gene ontology: tool for the unification of biology. The Gene Ontology Consortium, *Nat. Genet.*, 25:25–29, 2000.
- [4] Bates, G., Harper, P., and Jones, L., eds., Huntington's Disease, 3rd edn, Oxford University Press, 2002.
- [5] Becker, U. and Moldt, D., Object-oriented concepts for Coloured Petri nets, IEEE International Conference on Systems, Man and Cybernetics, 3:279–286, 1993.
- [6] Chen, M., Ona, V.O., Li, M., Ferrante, R.J., Fink, K.B., Zhu, S., Bian, J., Guo, L., Farrell, L.A., Hersch, S.M., Hobbs, W., Vonsattel, J.P., Cha, J.H., and Friedlander, R.M., Minocycline inhibits caspase-1 and caspase-3 expression and delays mortality in a transgenic mouse model of Huntington disease, *Nat. Med.*, 6:797–801, 2000.
- [7] Coleman, T.P., Tran, Q., and Roesser, J.R., Binding of a candidate splice regulator to a calcitoninspecific splice enhancer regulates calcitonin/CGRP pre-mRNA splicing, *Biochim. Biophys. Acta.*, 1625:153–164, 2003.
- [8] David, R. and Alla, H., Continuous Petri nets, In: 8th European Workshop on Application and Theory of Petri Nets, *Lecture Notes in Computer Science*, Springer-Verlag 340:275–294, 1987.
- [9] Doi, A., Nagasaki, M., Fujita, S., Matsuno, H., and Miyano, S., Genomic Object Net:II. Modeling biopathways by hybrid functional Petri net with extension, *Applied Bioinformatics*, 2:185–188, 2004.
- [10] Drath, R., Hybrid object nets: An object oriented concept for modeling complex hybrid systems, In: 3rd International Conference on Automation of Mixed Processes: Hybrid Dynamical Systems (ADPM), Shaker Verlag, 437–442, 1998.
- [11] Drath, R., Engmann, U., and Schwuchow, S., Hybrid aspects of modeling manufacturing systems using modified Petri nets, In: 5th Workshop on Intelligent Manufacturing Systems, 1999, http: //www.systemtechnik.tu-ilmenau.de/~drath/Download/Brasil98.ps.zip

- [12] Hackam, A.S., Singaraja, R., Wellington, C.L., Metzler, M., McCutcheon, K., Zhang, T., Kalchman, M., and Hayden, M.R., The influence of huntingtin protein size on nuclear localization and cellular toxicity, J. Cell. Biol., 141:1097–1105, 1998.
- [13] Hickey, M.A.and Chesselet, M.F., Apoptosis in Huntington's disease, Prog Neuropsychopharmacol Biol Psychiatry, 27:255–265, 2003.
- [14] Jacks, T., Power, M.D., Masiarz, F.R., Luciw, P.A., Barr, P.J., and Varmus, H.E., Characterization of ribosomal frameshifting in HIV-1 gag-pol expression, Nature, 331:280–283 1988.
- [15] Kim, Y.J., Yi, Y., Sapp, E., Wang, Y., Cuiffo, B., Kegel, K.B., Qin, Z.H., Aronin, N., and DiFiglia, M., Caspase 3-cleaved N-terminal fragments of wild-type and mutant huntingtin are present in normal and Huntington's disease brains, associate with membranes, and undergo calpaindependent proteolysis, *Proc. Natl. Acad. Sci. USA*, 98:12784–12789, 2001.
- [16] Kohn, K.W., Molecular interaction map of the mammalian cell cycle control and DNA repair systems, *Mol. Biol. Cell.*, 10:2703–2734, 1999.
- [17] Kuipers, B.J. and Shults, B., Reasoning in logic about continuous systems, Principles of Knowledge Representation and Reasoning: Proceedings of the Fourth International Conference KR94, Morgan Kaufmann 391–402, 1994.
- [18] Kummer, O., Introduction to Petri Nets and Reference Nets, Sozionik Aktuell, 1:1–9, 2001.
- [19] Maier, C. and Moldt, D., Object Coloured Petri Nets A Formal Technique for Object Oriented Modeling, In: Concurrent Object-Oriented Programming and Petri Nets, Advances in Petri Nets. *Lecture Notes in Computer Science*, Springer-Verlag, 2001:406–427, 2001.
- [20] Martin, J.B. and Gusella, J.F., Huntington's disease. Pathogenesis and management, N. Engl. J. Med., 315:1267–1276, 1986.
- [21] Matsuno, H., Tanaka, Y., Aoshima, H., Doi, A., Matsui, M., Miyano, S., Biopathways Representation and Simulation on Hybrid Functional Petri Net, In Silico Biol., 3:389-404, 2003. http://www.bioinfo.de/isb/toc_vol_03.html/
- [22] Matsuno, H., Doi, A., Nagasaki, M., and Miyano, S., Hybrid Petri net representation of gene regulatory network, *Pacific Symposium on Biocomputing 2000*, 5:341–352, 2000.
- [23] Matsuno, H., Murakami, R., Yamane, R., Yamasaki, N., Fujita, S., Yoshimori, H., and Miyano, S., Boundary formation by notch signaling in *Drosophila* multicellular systems: Experimental observations and gene network modeling by Genomic Object Net, *Pacific Symposium on Biocomputing* 2003, 8:152–163, 2003.
- [24] Matsuoka, S., Sarai, N., Kuratomi, S., Ono, K., and Noma, A., Role of individual ionic current systems in ventricular cells hypothesized by a model study, *The Japanese Journal of Physiology*, 53:105-123, 2003.
- [25] McCarthy, J., Recursive Functions of Symbolic Expressions and Their Computation by Machine, Part I, ACM Conference on Symbolic Manipulation, 3:184–195, 1960.
- [26] Mendes, P., GEPASI: A software package for modelling the dynamics, steady states and control of biochemical and other systems, *Comput. Appl. Biosci.*, 9:563–571, 1993.
- [27] Nagasaki, M., Doi, A., Matsuno, H., and Miyano, S., Genomic Object Net:I. A platform for modeling and simulating biopathways, *Applied Bioinformatics*, 2:181–184, 2004.
- [28] Nagasaki, M., Onami, S., Miyano, S., and Kitano, H., Bio-calculus: Its concept and molecular interaction, *Genome Informatics*, 10:133–143, 1999.
- [29] Reddy, V.N., Liebman, M.N., and Mavrovouniotis, M.L., Qualitative analysis of biochemical reaction systems, *Comput. Biol. Med.*, 26:9–24, 1996.

- [30] Reddy, V.N., Mavrovouniotis, M.L., Liebman, M.N., Petri Net Representations in Metabolic Pathways, In Proceedings First International Conference on Intelligent Systems for Molecular Biology, Volume 1., MIT Press, 328–336, 1993.
- [31] Reisig, W. and Rozenberg, G., eds., Lecture on Petri nets I: Basic models, Lecture Notes in Computer Science, Springer-Verlag, 1491, 1998.
- [32] Roy, S., Bayly, C.I., Gareau, Y., Houtzager, V.M., Kargman, S., Keen, S.L., Rowland, K., Seiden, I.M., Thornberry, N.A., and Nicholson, D.W., Maintenance of caspase-3 proenzyme dormancy by an intrinsic "safety catch" regulatory tripeptide, *Proc. Natl. Acad. Sci. USA*, 98:6132–6137, 2001.
- [33] Roizin, L., Stellar, S., and Liu, J.C., Neuronal nuclear-cytoplasmic changes in Huntington's chorea: electron microscope investigations, *Raven Press*, 1979.
- [34] Senut, M.C., Suhr, S.T., Kaspar, B., and Gage, F.H., Intraneuronal aggregate formation and cell death after viral expression of expanded polyglutamine tracts in the adult rat brain, J. Neurosci., 20:219–229, 2000.
- [35] Tomita, M., Hashimoto, K., Takahashi, K., Shimizu, T.S., Matsuzaki, Y., Miyoshi, F., Saito, K., Tanida, S., Yugi, K., Venter, J.C., and Hutchison, C.A.3rd., E-CELL: software environment for whole-cell simulation, *Bioinformatics*, 15:72–84, 1999.
- [36] Tran, Q., Coleman, T.P., and Roesser, J.R., Human transformer 2beta and SRp55 interact with a calcitonin-specific splice enhancer, *Biochim. Biophys. Acta.*, 1625:141–152, 2003.
- [37] The Huntington's Disease Collaborative Research Group, A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes, *Cell*, 72:971–83, 1993.
- [38] Webster, S.D., Cason, Z., Lemos, L.B., and Benghuzzi, H., Cytohistologic correlation in patients with clinical symptoms of postmenopausal bleeding, *Biomed. Sci. Instrum.*, 36:367–372, 2000.
- [39] Wheeler, V.C., White, J.K., Gutekunst, C.A., Vrbanac, V., Weaver, M., Li, X.J., Li, S.H., Yi, H., Vonsattel, J.P., Gusella, J.F., Hersch, S., Auerbach, W., Joyner, A.L., and MacDonald, M.E., Long glutamine tracts cause nuclear localization of a novel form of huntingtin in medium spiny striatal neurons in HdhQ92 and HdhQ111 knock-in mice, *Hum. Mol. Genet.*, 9:503–513, 2000.
- [40] Wellington, C.L., Ellerby, L.M., Gutekunst, C.A., Rogers, D., Warby, S., Graham, R.K., Loubser, O., van R., J., Singaraja, R., Yang, Y.Z., Gafni, J., Bredesen, D., Hersch, S.M., Leavitt, B.R., Roy, S., Nicholson, D.W., and Hayden, M.R., Caspase cleavage of mutant huntingtin precedes neurodegeneration in Huntington's disease, *J. Neurosci.*, 22:7862–7872, 2002.
- [41] Wellington, C.L., Ellerby, L.M., Hackam, A.S., Margolis, R.L., Trifiro, M.A., Singaraja, R., McCutcheon, K., Salvesen, G.S., Propp, S.S., Bromm, M., Rowland, K.J., Zhang, T., Rasper, D., Roy, S., Thornberry, N., Pinsky, L., Kakizuka, A., Ross, C.A., Nicholson, D.W., Bredesen, D.E., and Hayden, M.R., Caspase cleavage of gene products associated with triplet expansion disorders generates truncated fragments containing the polyglutamine tract, J. Biol. Chem., 273:9158–9167, 1998.
- [42] GON: Genomic Object Net project, http://GenomicObject.net/
- [43] HFPNe Generic Models, http://genomicobject.net/public/BSB2004/code/
- [44] Pnuts, http://www.pnuts.org/