

1 Development of microsatellite markers in cultivated and wild species of sections *Cepa*
2 and *Phyllodolon* in *Allium*

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1 **Abstract** The potential of microsatellite markers for use in genetic studies has
2 been evaluated in *Allium* cultivated species (*Allium cepa*, *A. fistulosum*) and its allied
3 species (*A. altaicum*, *A. galanthum*, *A. oscaninii*, *A. roylei*, *A. vavilovii*). A total of 77
4 polymerase chain reaction (PCR) primer pairs were employed, 76 of which amplified a
5 single product or several products in either of the species. The 29 AMS primer pairs
6 derived from *A. cepa* and 47 microsatellites primer pairs from *A. fistulosum* revealed a
7 lot of polymorphic amplicons between seven *Allium* species. Some of the microsatellite
8 markers were effective not only for identifying an intraspecific F₁ hybrid between
9 shallot and bulbonion but also for applying to segregation analyses in its F₂ population.
10 All of the microsatellite markers can be used for interspecific taxonomic analyses
11 among two cultivated and four wild species of sections *Cepa* and *Phyllodolon* in *Allium*.
12 Generally, our data support the results obtained from recently performed analyses using
13 molecular and morphological markers. However, the phylogeny of *A. roylei*, a
14 threatened species with several favorable genes, was still ambiguous due to its different
15 positions in each dendrogram generated from the two primer sets originated from *A.*
16 *cepa* and *A. fistulosum*.

17 **Keywords** Allium, microsatellite markers, DNA polymorphism

18

1 **Introduction**

2 The abundance, characterization, and the usefulness of SSR markers have been
3 reviewed in cultivated *Allium* species (McCallum, 2007). A number of microsatellites
4 have been identified and characterized in bulb onion, *Allium cepa* Common onion group
5 (Fischer and Bachmann 2000) and bunching onion, *Allium fistulosum* (Song et al. 2004;
6 Tsukazaki et al. 2007). The previous studies implied that such microsatellites could be
7 applied as markers for several kinds of genetic analyses in a wide range of species in
8 sections *Cepa* and *Phyllodolon* of *Allium*. In fact, a number of bunching onion
9 microsatellites could be applicable for the purity determination of F₁ seeds as well as
10 variety identification (Tsukazaki et al. 2006). Tsukazaki et al. (2009) demonstrated an
11 SSR-tagged breeding scheme for *A. fistulosum*. Furthermore, many bulb onion
12 microsatellites flanking primer pairs proved to be functional not only in shallot (*A. cepa*
13 *Aggregatum* group) but also in bunching onion (Masuzaki et al. 2006). On the other
14 hand, section *Cepa* of *Allium* includes several wild species, i.e. *Allium galanthum*,
15 *Allium oschaninii*, *Allium vavilovii*, etc. as well as the two cultivated crops (bulb onion,
16 shallot) (Vvedensky 1944; Havey 1995). With two wild species, *Allium altaicum* and
17 *Allium microbulbum*, another cultivated crop, bunching onion was placed at section
18 *Phyllodolon* (Vvedensky 1944). Bulb onion microsatellites had been applied for

1 assessing interspecific relatedness within these species (Fischer and Bachmann 2000).
2 Some of the microsatellite markers can be used for interspecific taxonomic analyses
3 among close relatives of bulb onion. There has this far been no example applied to other
4 species by extending bunching onion microsatellites.

5 *Allium roylei* has been known as a potential gene reservoir for introducing
6 several disease resistances to bulb onions (Kik 2002) and shows a quite unique position
7 in the taxonomy of genus *Allium* (Fritsch and Friesen 2002). This wild species crosses
8 easily with *A. cepa* and *A. fistulosum*, and shares a high degree of genetic similarity
9 with other species in the two sections mentioned above. Nonetheless, most
10 morphological characters differ remarkably from others in these sections and are much
11 more similar to those of section *Oreiprason* (Fritsch and Friesen 2002). Further recent
12 evidence indicates that *A. roylei* might have a hybrid origin, as its nuclear DNA profile
13 is related to species of the sections *Cepa* and *Phyllodolon* but its chloroplast DNA
14 profile to the section *Schoenoprasum* (van Raamsdonk et al. 1997, 2000).

15 In the present study, the microsatellite marker analyses using several species
16 and their hybrids were performed with the following objectives: (1) development of
17 microsatellite markers useful for marker-assisted selection approach using alien genes in
18 the cultivated species of sections *Cepa* and *Phyllodolon*; (2) evaluation of heritability

1 via the simple test of a F₁ hybrid and the segregation analysis of its F₂ population; (3)
2 verification of genome organization in *A. roylei* which is an insufficiently known wild
3 species in its phylogenesis.

4

5 **Materials and Methods**

6 **Plant materials**

7 Doubled haploid line of shallot (*Allium cepa* L. Aggregatum group, 'DHA',
8 genomes AA), bulb onion (*A. cepa* L. Common onion group, 'Hayatama-2', CC), an F₁
9 intraspecific hybrid between shallot and bulb onion ('DHA' × 'Hayatama-1', AC),
10 Japanese bunching onion (*Allium fistulosum* L., 'Kujo-hoso', FF), *Allium altaicum* Pall.
11 (I: '97010-25', II: '97213-15'), *Allium galanthum* Kar. et Kir. (I: '97205-22', II:
12 '97205-42'), *Allium roylei* Stearn (I: '95001-2', II: '95001-6'), and *Allium vavilovii* M.
13 Pop. et Vved. (I: '97202-8', II: '97202-2') were used for detecting intra- and
14 interspecific polymorphisms. 'DHA' was obtained via the chromosome doubling of a
15 gynogenetic haploid from diploid shallot 'Chiang Mai'. Its F₂ population with 38 plants
16 was employed to examine allelic relationships at some SSR loci.

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1 DNA extraction and PCR amplification

2 Total genomic DNA was isolated from fresh leaf tissue using a mini-prep
3 DNA-isolation method (van Heusden et al. 2000) except for shallot, bulb onion, and
4 their F₁ intraspecific hybrids, whose total genomic DNAs were extracted from the
5 leaves according to the procedure of Dubouzet et al. (1996). Thirty microsatellite primer
6 sets (AMS01 to AMS30; GenBank accession numbers, AJ391666 to AJ391725) for bulb
7 onion developed by Fischer and Backmann (2000), 46 microsatellite primer pairs
8 (bunching onion microsatellite, BMS) derived from SSR-enriched DNA libraries of
9 Japanese bunching onion reported by Wako et al. (2002), Song et al. (2004), and
10 Tsukazaki et al. (2007), and one onion EST-derived SSR primer pairs (Kuhl et al. 2004)
11 were examined. PCR amplifications for the AMS primer sets were carried out according
12 to the procedure of Masuzaki et al. (2006). For the microsatellite primer sets from
13 bunching onion, PCR mixture was made with 0.8 μM of each of the primers, 0.2 mM
14 dNTPs, 1 × *rTaq* buffer, 0.5 U *rTaq* polymerase (Takara, Shiga, Japan), and 100 ng
15 template DNA in a volume of 20 μL. PCR temperatures were performed with the minor
16 modifications of the procedure of Ohara et al. (2005). PCRs were done in a program
17 thermal cycler (iCycler; Bio-Rad, Hercules, USA), in which the ramp times were
18 carried out in the default conditions that adjusted temperatures at the maximum ramp

1 rate with the minimum ramp time. PCR products were electrophoresed and separated on
2 2% (w/v) agarose gel. If the PCR products were monomorphic on agarose gel, they
3 were subjected to denaturing polyacrylamide gel electrophoresis (PAGE) with silver
4 staining. The amplified fragments from 100 to 700 bp were scored in each *Allium*
5 species. The markers were designated according to the primer name and the molecular
6 size of each band in base pairs.

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8 Phylogenetic analysis

9 The microsatellite fragments obtained in seven species expect for F₁
10 intraspecific hybrids between shallot and bulb onion were scored as either present or
11 absent. The genetic distances were calculated according to the procedure of Nei and Li
12 (1979) by using a program Restdist (Program to compute distance matrix from
13 restriction sites or fragments) in software PHYLIP, version 3.68 (Felsenstein 1989).
14 Phylogenetic analyses were performed using the Neighbor Joining Method (Saitou and
15 Nei 1987). The graphic output of the phylogenetic trees was created with the MEGA
16 version 4 program (Tamura et al. 2007)

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1 **Results**

2 Polymorphisms between *Allium* species

3 The 29 AMS primer pairs, in which the primer pairs for AMS11 amplified no
4 bands in all the investigated plants, and the 47 bunching onion microsatellites, or BMS,
5 primer pairs, (including one onion EST-SSR primer pair, ACM071) revealed a lot of
6 polymorphic amplicons between seven *Allium* species (Figs. 1 and 2; Suppl. Tables 1
7 and 2). A total of 782 bands were obtained by the 28 AMS primer pairs, and a total of
8 393 bands by 47 BMS primer pairs. The microsatellite markers of *A. fistulosum* were
9 obtained in 26 AMS and 46 BMS primer pairs; bulb onion, 28 and 24; shallot, 28 and
10 28; *A. altaicum*, 26 in I / 24 in II and 38 in I / 43 in II; *A. galanthum*, 25 in I / 22 in II
11 and 26 in I / 15 in II; *A. roylei*, 26 in I / 25 in II and 19 in I / 23 in II; *A. vavilovii*, 28 in I
12 / II and 17 in I / 13 in II (Suppl. Tables 1 and 2). The average number \pm standard
13 deviation of polymorphic bands per AMS and bunching onion microsatellites primer
14 were, respectively, 26.1 ± 25.8 (0 in AMS11 to 125 in AMS05) and 8.4 ± 4.9 (1 in three
15 primer sets to 21 in AFC02E08) across the investigated plants (Fig. 2).

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1 Evaluation of heritability of microsatellite markers in F₁ and F₂ between shallot and
2 onion

3 Twenty-eight AMS primer pairs except for AMS11 and AMS24 amplified the
4 fragments in F₁ intraspecific hybrid, which generated by crossing bulb onion with
5 shallot, as well as the two species. Of the 28 AMS primer pairs, some microsatellite
6 markers yielded in AMS03, AMS17, and AMS21 exhibited significant heritabilities on
7 denaturing PAGE with silver staining (Fig. 3a, Table 1). The segregation of these
8 markers was found to fit the monogenic segregation of 1 : 2 : 1 (Fig. 4). In addition,
9 amplicons were observed in 23 out of the 47 bunching onion microsatellites primer
10 pairs. Of the 23 primer pairs, only ACE127 produced heritable two microsatellite
11 markers, i.e. ACE127-260 and -276 in *A. cepa* (Fig. 3b).

12 Simultaneous segregations at three loci were observed as shown in Table 2.
13 Three different simultaneous segregations were tested for independence by chi-square
14 tests. None of the data showed a significant difference from the expectation of
15 independent segregation (Table 2), suggesting that these three loci are located on
16 different chromosomes or distantly on the same chromosome. The chromosomal
17 location of the AMS17 locus could be understood since one marker, AMS17-270, at
18 same locus was already assigned to the chromosome 1 of *A. cepa* in our previous study

1 (Masuzaki et al. 2006).

2

3 Verification of genome organization in *Allium roylei*

4 The relationships among the entries are shown by the phyllograms in Fig. 4 on
5 the basis of genetic distances calculated by using polymorphic amplicons between the
6 seven *Allium* species (143 sites for AMS primer pairs and 382 sites for BMS primer
7 pairs). The phyllogram of BMS primer pairs (Fig. 4b) showed two major nodes
8 consisting of (I) the three species of the section *Cepa* (*A. vavilovii*, *A. cepa*, *A.*
9 *galanthum*) and *A. roylei* as well as (II) the two species of the section *Phyllodolon* (*A.*
10 *fistulosum* and *A. altaicum*), in agreement with one promising previous report (van
11 Raamsdonk et al 2000). However, the results of the AMS primer pairs were different
12 from the ones of the BMS primer pairs in clustering of *A. roylei*. While, *A. roylei* was
13 clustered in one neutral node together with *A. galanthum* out of three different nodes
14 (Fig. 4a). Other two nodes consisted separately of the sections *Cepa* and *Phyllodolon*.

15

16 **Discussion**

17 The most conspicuous feature of *Allium* nuclear genomes is their great size,
18 which has made *Allium* crops complicates most molecular genetic analysis. For instance,

1 bulb onion nuclear genome contains 17.9 pg or 15,290 Mbp of DNA per 1C nucleus,
2 and making it one of the hugest genomes among cultivated plants (Havey 2002).
3 Bunching onion shows a 28 % smaller genome size than bulb onion (Ricroch et al.
4 2005). There seems to be a big difference between these species in not only genome size
5 but also its structure. Jones and Rees (1968) and Narayan (1988) observed that all eight
6 bivalents were asymmetric in interspecific hybrids between the two species. This
7 indicated that genomic DNA differences were spread across all eight chromosomes.
8 Chiasmata localized near the centromere of bunching onion could be explained if this
9 region were largely euchromatic and therefore gene-rich (Havey 2002). The integration
10 of linkage and physical maps in bunching onion showed that recombination
11 predominantly occurs in the proximal half of chromosome arms and that 57.9% of
12 *PstI/MseI* AFLP markers are located in close proximity to the centromeric region
13 (Khrustaleva et al. 2005). These results suggested the presence of genes in this region.
14 Tsukazaki et al. (2008) constructed a chromosome-specific linkage map of bunching
15 onion based on the SSR markers used in this study. Linkage groups on same
16 chromosome were split into two parties except a few cases, suggesting the presence of
17 repetitive sequences (e.g. microsatellites) in the distal half of each chromosome arms.
18 On the other hand, in bulb onion two different medium-density intraspecific and

1 interspecific linkage maps have thus far been developed based on several sorts of DNA
2 markers (RFLP, SSR, SNP, InDel) (Martin et al. 2005) and *EcoR1/Mse1* AFLP markers
3 (van Heusden et al. 2000), respectively. The integration of linkage map from two
4 different species would be possible by plotting a number of common codominant
5 markers, such as our promising microsatellite markers with heritability and leads us to
6 reveal a trace of the chromosomal structural changes occurred in their speciation
7 process.

8 *Allium roylei* seems to be a potential gene reservoir for bunching onion as well
9 as for bulb onion (Kik 2002). This wild species shows combined resistant to Botrytis
10 leaf blight (*Botrytis squamosa*) (Lacy and Lorbeer 2008) and downy mildew
11 (*Peronospora destructor*) (Schwartz 2008) in bulb onion. For the downy mildew, a
12 resistance locus had been anchored to the distal end of chromosome 3R in *A. roylei* and
13 then was successfully introduced into a bulb onion cultivar via the use of a conventional
14 backcrossing procedure (Scholten et al. 2007). Nowadays many peoples in *Allium*
15 research community are very interested in the origin or taxonomic position of *A. roylei*.
16 However, one *A. roylei* strain was introduced into the European research scene in the
17 1960s. All living plants investigated in the world are derived from this single fertile
18 strain (Fritsch and Friesen 2002). Sharma and Gohil (2003) had recently exploited a

1 small threatened population of *A. roylei* from the Bani region of Jammu province in
2 India and observed the meiosis of PMCs in ten plants equally from two populations of *A.*
3 *roylei*. The results revealed multivalent associations involving the entire complements
4 of 16 chromosomes. The subsequent observation of somatic chromosomes in same
5 populations clarified the extensive karyotypic variability due to repeated interchanges
6 involving a number of non-homologous chromosomes (Sharma and Gohil 2003). The
7 evaluation of its genetic diversity by microsatellite polymorphisms would provide
8 informative parameters not only to carry out a hazard prediction for its threatened status
9 but also to enforce both *in situ* and *ex situ* conservation strategies for this threatened
10 species.

11

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Table 1. Segregation patterns of three AMS primers in F₂ progeny

Primer ^z (shallot/onion)	Genotype observed ^y				χ^2	Probability in χ^2 -test for 1 : 2 : 1
	<i>a</i>	: <i>ab</i>	: <i>b</i>	obscurity		
AMS03 (196/204)	8	19	9	2	0.263	.900-.750
AMS17 (282/268)	9	18	9	2	0.105	.950-.900
AMS21 (284/277)	8	19	11	0	0.476	.900-.750

^z Arabic numerals in parenthesis show fragment size.

^yAllele *a* is derived from shallot, and *b* from bulb onion.

Table 2. Estimation of linkage between microsatellite loci derived from AMS primers

Loci tested for linkage	Type of freq.	Observed numbers and expected numbers (below) of genotypes										No. of plants	χ^2 -test for independence	d.f.	Prob.
		aa.aa	aa.ab	aa.bb	ab.aa	ab.ab	ab.bb	bb.aa	bb.ab	bb.bb	obscurity				
AMS17/AMS03	Obs.	1	6	2	2	8	6	5	3	1	4	34	7.941	4	.100-.050
	Exp.	2.125	4.25	2.125	4.25	8.5	4.25	2.125	4.25	2.125					
AMS17/AMS21	Obs.	2	5	2	4	8	6	1	5	3	2	36	0.889	4	.950-.900
	Exp.	2.25	4.5	2.25	4.5	9	4.5	2.25	4.5	2.25					
AMS03/AMS21	Obs.	3	3	3	3	13	2	2	2	5	2	36	9.222	4	.100-.050
	Exp.	2.25	4.5	2.25	4.5	9	4.5	2.25	4.5	2.25					

Figure legends

Figure 1. Amplification profiles of the primer set for AMS03 (**a**) and AFB02C01 (**b**) in *Allium* species. M, molecular size marker (100 bp DNA Ladder); CC, bulb onion; AA, shallot; FF, *A. fistulosum*; RR, *A. roylei*; GG, *A. galanthum*; LL, *A. altaicum*; VV, *A. vavilovii*.

Figure 2. Number distribution of 30 AMS primer sets (■), and 46 primer sets (bunching onion microsatellite, BMS) derived from SSR-enriched DNA libraries of *Allium fistulosum* and one onion EST-derived SSR primer pair (□) among the number of microsatellite markers.

Figure 3. Evaluation of heritability for microsatellite markers (**a**, AMS03-196, -204 and -214; **b**, ACE127-240, -260 and -276) in F₁ hybrid between shallot and bulb onion. M, molecular size marker (100 bp DNA Ladder); AA, shallot; CC, bulb onion; AC, F₁ intraspecific hybrid between shallot and bulb onion.

Figure 4. Segregation of F₂ population which originated from a single F₁ plant between shallot and bulb onion. S, shallot; B, Bulb onion; M, molecular size marker (100 bp DNA Ladder). An upper arrow indicates AMS03-204. A lower arrow indicates AMS03-196.

Figure 5. Phylograms generated by Neighbor Joining Method analyses of microsatellite markers amplified using 30 AMS primer sets (**a**), and 46 BMS primer sets (**b**) in seven *Allium* species. A scale of genetic distances is shown.

Figure 1, Araki et al.

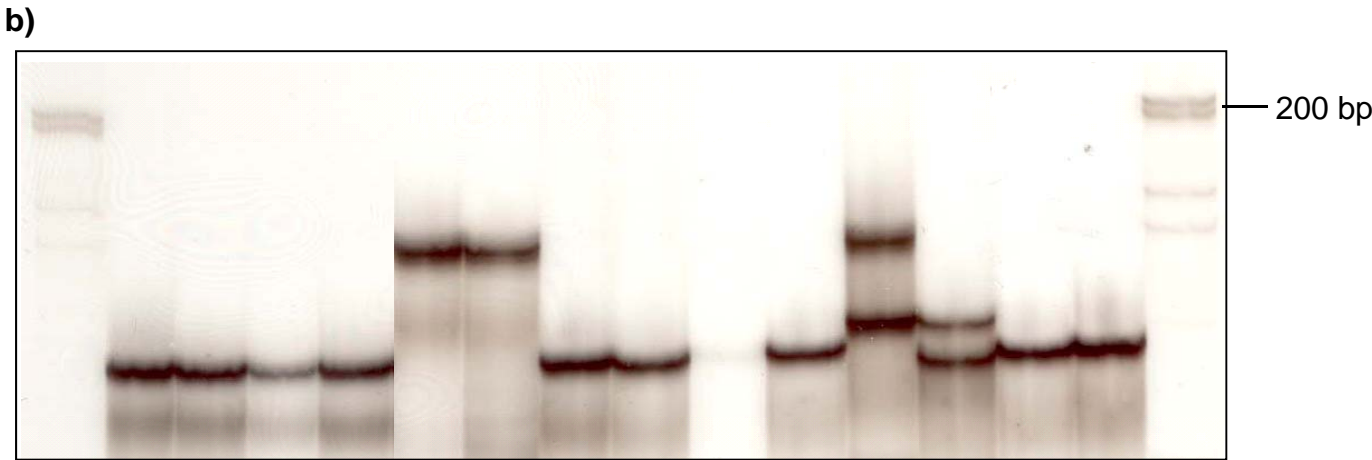
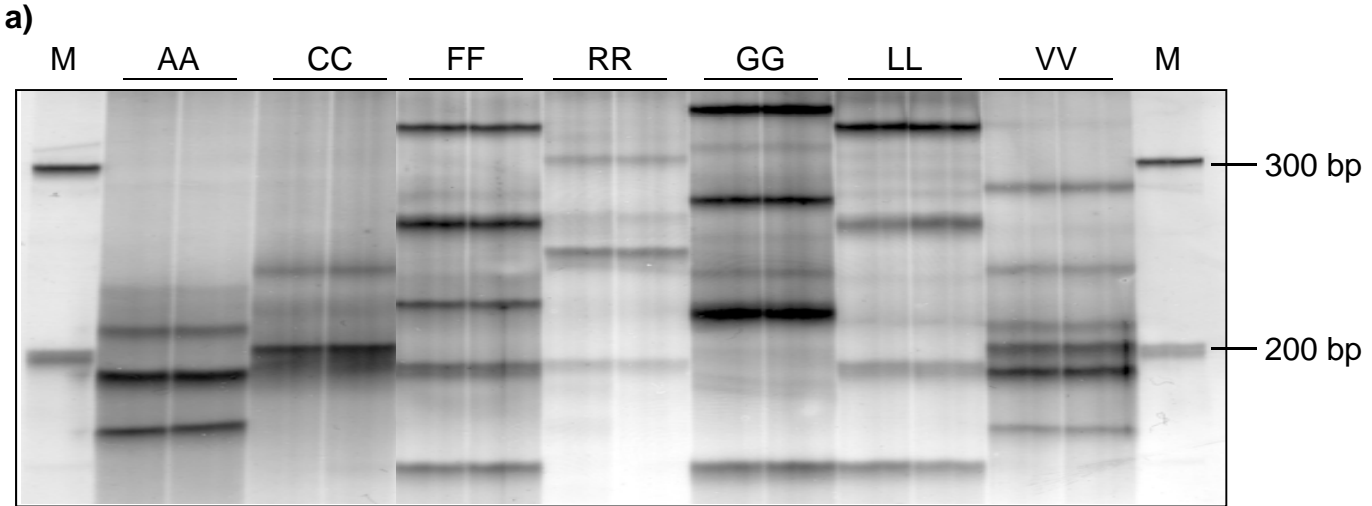


Figure 2, Araki et al.

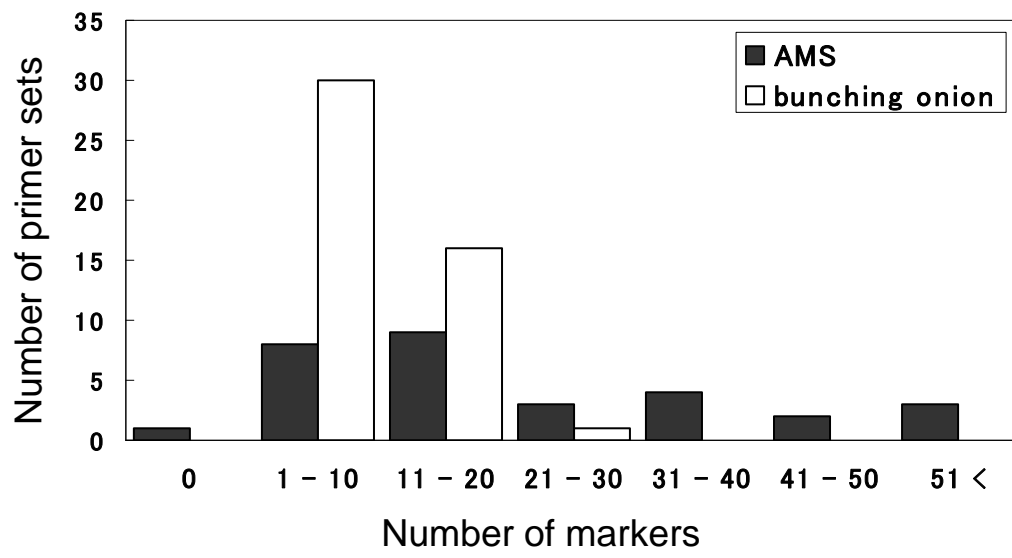


Figure 3, Araki et al.

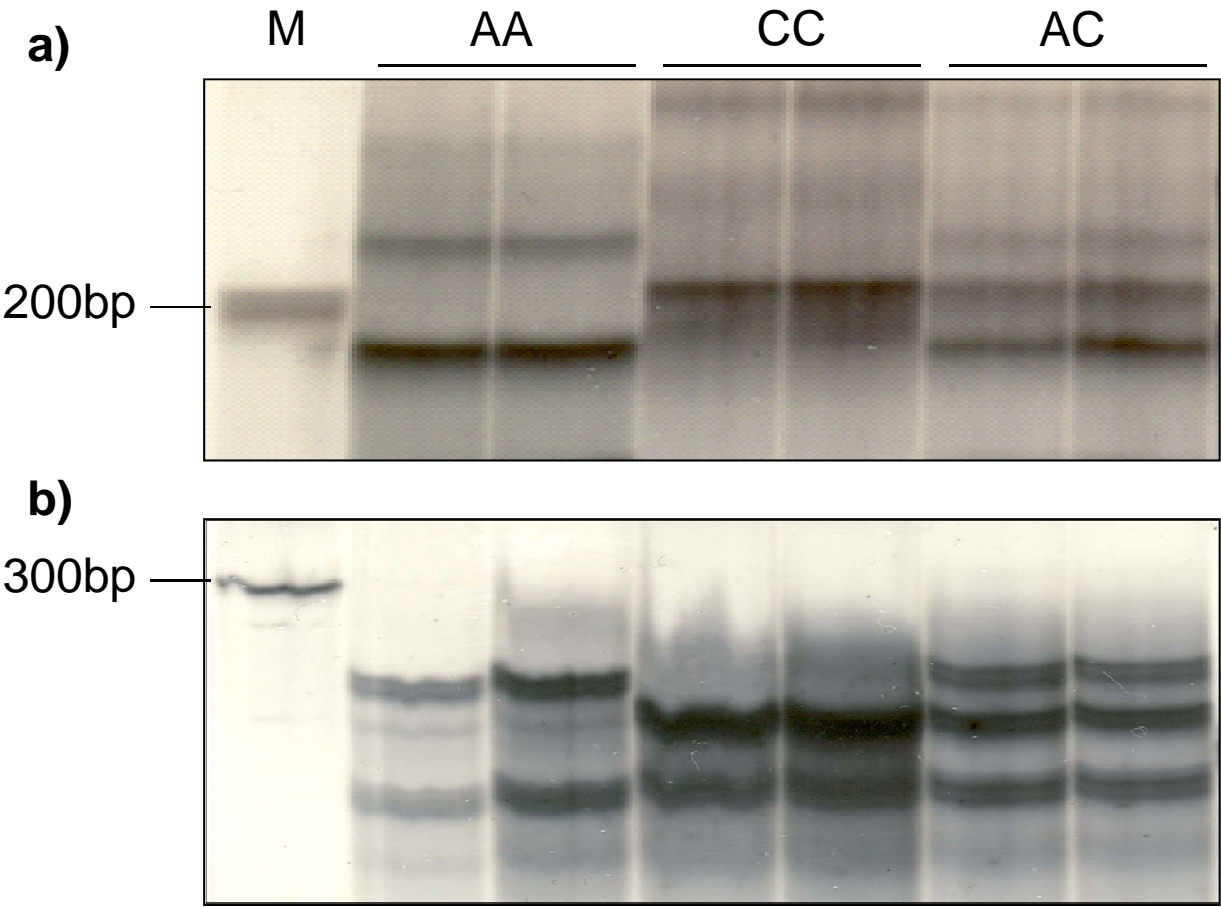


Figure 4, Araki et al.

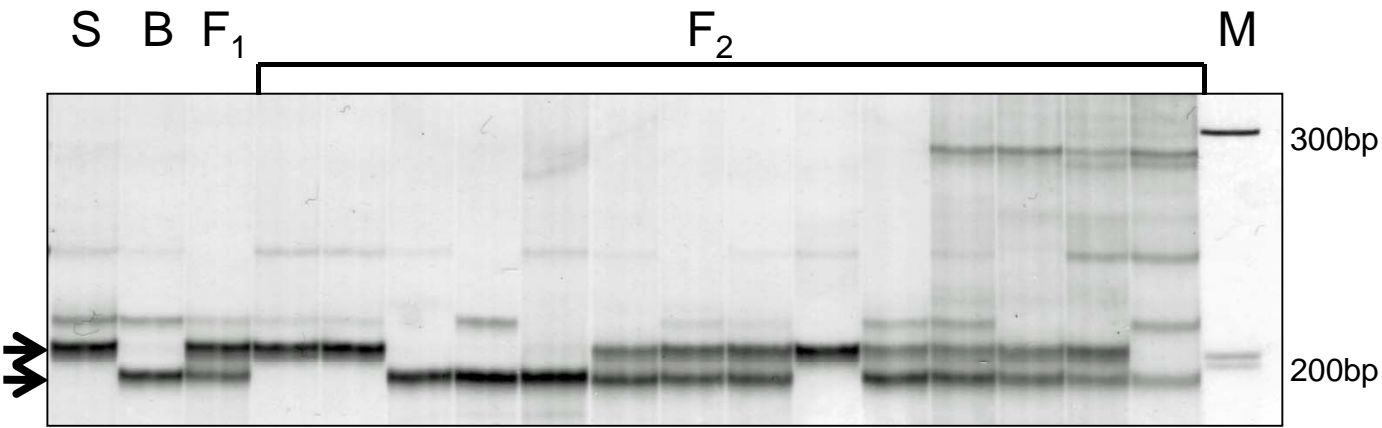
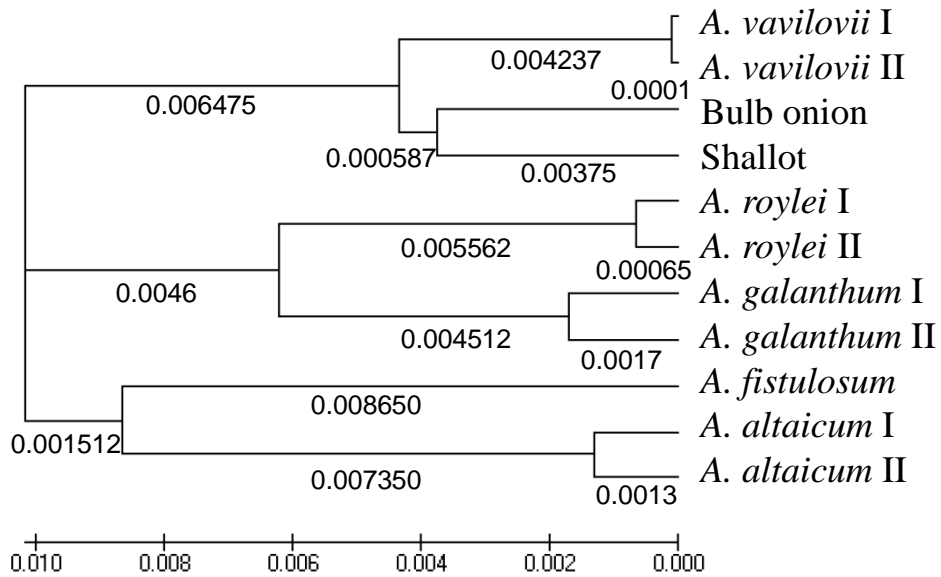
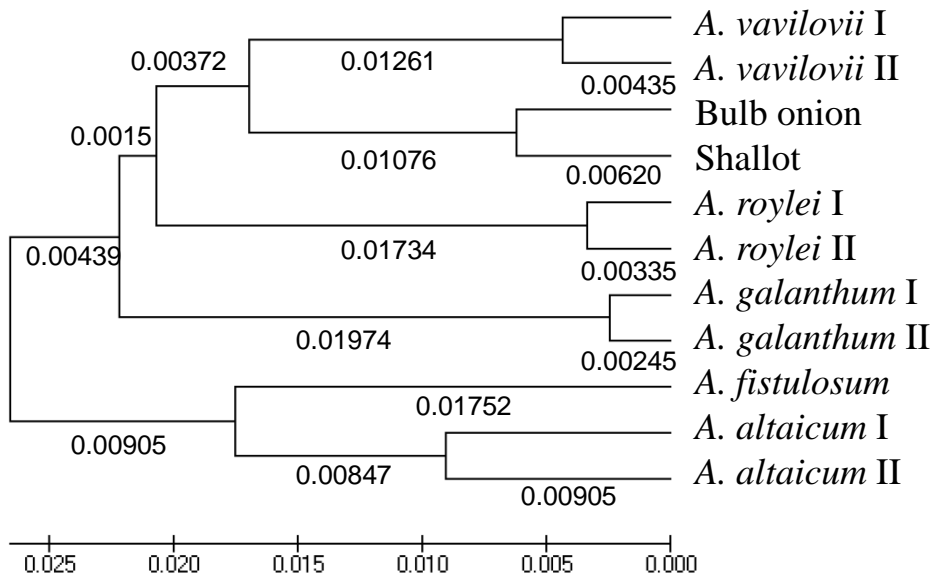


Figure 5, Araki et al.

a)



b)



06	-		216, 289	216, 289	216, 286	-	-		209	209		209, 279	253, 279	209, 279	253, 279	216, 289	216, 289			
07	109, 175, 567	121, 293, 175,	109, 121, 175,	109, 121, 175,	109, 121, 175,	109, 175, 567	121, 293, 567	109, 175, 567	121, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447	109, 121, 175, 293, 351, 447
08	181, 337, 497, 538, 563, 669	328, 356, 508, 553, 655,	196, 209, 216, 356, 638	196, 198, 356	196, 198, 356	181	-	-	-			457, 634	617, 356			196, 198	196, 198			
09	222, 240		240, 260	222, 240, 259, 268, 482, 631	240, 260, 259, 280, 393	222, 240, 259, 406, 542	240, 393, 259	-		240, 406	393, 625	240, 393, 625	265, 416, 393	240, 265, 393	265, 393, 406	240, 275, 393, 406	240, 406	240, 393,		
10	125, 149, 164, 228,	145, 161, 216, 256,	145, 149, 151, 156,	145, 149, 151, 156,	145, 149, 151, 156,	226, 256, 357, 538,	228, 265, 365, 560,	226, 256, 357, 538,	228, 265, 365, 560,	145, 226, 256, 491,	149, 228, 265, 505,	145, 228, 265, 505,	149, 216, 228, 265, 268,	145, 226, 216, 228, 256, 268,	149, 226, 216, 228, 256, 268,	145, 153, 170, 293,	149, 166, 284, 448,	145, 153, 170, 293,	149, 166, 284, 448,	

			600	447,	500,																
				463,	522,																
				477,	597																
				492,																	
				500,																	
				550,																	
				597,																	
				600,																	
				627,																	
				650,																	
				668																	
16	116,	217,	129,	129,	129,	116,	223,	116,	223,	113,	119,	113,	119,	119,	124,	119,	124,	129,	142,	129,	142,
	223,	234,	140,	142,	134,	230,	239,	230,	239,	126,	134,	126,	134,	136,	140,	136,	140,	165,	168,	165,	168,
	258,	281,	168,	168,	142,	243,	258,	243,	258,	140,	168,	140,	168,	234,	242,	234,	242,	179,	181,	179,	181,
	441		169,	258,	146,	281		281		234,	239,	234,	239,	258		258		188,	230,	188,	230,
			179,	266,	154,					243,	258	243,	258					239,	258,	239,	258,
			185,	271	168,													266,	271,	266,	271,
			190,		244,													277		277	
			244,		258,																
			258,		266,																
			261,		271																
			277																		
17	459,	541,	109,	274,	184,	-		184,	187,	184,	187,	184,	187,	184,	187,	184,	187,	184,	187,	184,	187,
	559,	572	271,	282,	187,			192,	195,	190,	192,	190,	192,	195,	197,	195,	197,	268,	274,	268,	274,
			277,	360,	268,			556,	572,	195,	222,	195,	222,	222,	227,	222,	227,	360,	367	360,	367
			360,	367	274,			654		227,	509,	227,	509,	334,	343,	334,	343,				

				473,																	
				617,																	
				629																	
24	154,	179,	-	-	-	154,	179,	154,	179,	424,	736,	424,	736,	-	-	-	-				
	396,	736,				398,	736,	398,	736,	745		745									
	745					745		745													
25	169,	178,	120,	116,	120,	178,	200,	178,	200,	153,	169,	153,	169,	169,	178,	169,	178,	206,	218,	206,	218,
	205,213		178,	120,	178,	205,213		205,213		178,	205,	178,	205,	205,	213,	205,	213,	222		222	
			206,	121,	206,					213		213		266		266					
			218,	178,	218,																
			225,	206,	225,																
			258,	218,	235,																
			270	225,	243																
				235,																	
				270																	
26	609,	614,	179,	179,	179,	262,274	-			202,	205,	202,205		202,205		202,205		179,	181,	179,	181,
	657		181,	181,	181,					558								205,208		205,208	
			189,	205,	189,																
			205,	208	205,																
			208		208																
27	234,	278,	211,	211,	308,	333,	395,	333,	395,	265,	347,	265,	333,	265,	288,	265,	288,	268,	308,	268,	308,
	294,	308,	214,	214,	320,	408,	443,	408,	443,	421,	573,	347,	377,	368,	421,	368,	421,	333,	347,	333,	362,
	331,	368,	234,	234,	421,	573		573		683		573,683		443,	573,	443,	573,	443,	463,	408,	439,
	391,	408,	265,	265,	463,									660		660		480,	500,	443,	463,
	421,	443,	281,	294,	480,													538,	627,	480,	500,
	510,	573,	300,	308,	538,													640		538,	627,

			540,														
			575,														
			603,														
			634,														
			672														
29	353, 694	293,	299,	293,	353,	488,	353,	488,	424, 694	424,	590,	175,	179,	175,	179,	353, 694	353, 694
		315,	330,	315,	490, 560		490, 560			596,	681,	600		600			
		327	353,	327						694							
			520,														
			540,														
			681,														
			694														
30	343, 353	343,	412,	343,	353, 607		353,	424,	499, 510	683, 697		325		325		343, 353	343, 353
		353	633	353			607										

^z The AMS numbers were described by Fischer and Bachmann (2000).

^y “-” indicates absence of amplified fragments in the species.

^x intraspecific hybrid between shallot and bulb onion

Suppl. Table 2. Fragment sizes amplified from *Allium* species by 46 microsatellite primer sets (bunching onion microsatellite, BMS) derived from SSR-enriched DNA libraries of *Allium fistulosum* developed by Tsukazaki et al. (2007) and onion EST-derived SSR primer pair (Kuhl et al., 2004).

SSR locus ^z	Observed size (bp) ^y												
	<i>A. fistulosum</i>		<i>A. cepa</i>		F ₁ ^x	<i>A. altaicum</i>		<i>A. galanthum</i>		<i>A. roylei</i>		<i>A. vavilovii</i>	
	onion		shallot			I'97010-25'	II'97213-15'	I'97205-22'	II'97205-42'	I'95001-2'	II'95001-6'	I'97202-8'	II'97202-2'
AFA06H07	243, 270		274	274	274	274	274	-	-	270	270	-	-
AFA03F08	264, 295		218, 241, 277, 291	218, 241, 273, 286	282, 291, 400	236, 259, 264	259, 264, 320	176, 218, 236, 259, 268	176, 218, 236, 259, 268	110, 170, 236, 241, 245, 250	-	-	-
AFB04B08	151, 170		229, 411	229, 411	229, 267	149, 168	147, 174	138, 147	138, 147	149	149	-	-
AFB05H03	222, 283		211, 283	211, 283	211, 283	272, 289	272, 300	-	283	-	-	-	-
AFA01A04	273		-	-	-	286	273	-	-	-	-	-	-
AFA01H03	117, 172, 175, 179	143	143	143	143	100, 121, 153, 183	158	145	145	143	143	145	145
AFS012	205, 209	-	183, 353, 363, 437	183, 353, 363, 437	198, 205	200, 205	196, 200	468	-	194, 198, 295, 321, 374, 416, 437, 458	-	-	-
AFAT04E09	282	-	-	-	-	-	255, 277	108, 223	-	-	179	-	-
AFB02C01	129, 173	144, 152	144, 152	144, 152	144, 152	119, 158, 175	112, 119, 152, 158	-	115, 154	110, 152	110, 152	112, 154	112, 154
AFRA04D09	164	-	-	-	-	164, 166	156, 160	471	-	-	274, 279	-	-
AFA01H12	190, 198	164, 192	154, 164	154, 164	-	164, 211	190, 198	-	-	-	168	-	-
AFS104-2	205, 225	-	174, 184, 192, 200	174, 184, 192, 200	-	210, 220, 225	170, 178, 188, 194	182	-	-	170, 176, 250	194, 205	-
AFA02A05	268, 286	-	282, 327	282, 327	-	-	277	-	-	-	273	-	-

AFC02E08	116, 168	166, 180, 184, 188	166, 180, 184, 188, 205, 225, 230, 265, 487, 600	180, 184, 188	166, 215, 245, 275	120, 168	162	162	100, 134, 148	100, 134, 148	122, 168	122, 168
AFS149	194, 205	232, 253	226	226, 253	190, 198	190, 200, 216	181, 190	181, 190	185, 194	185, 194	253	253
AFA01F03	136, 138, 145, 147, 224, 238	-	-	-	173, 187, 233	140, 149, 167, 178, 223	-	-	-	-	233	233
AFS042	150, 157, 243, 248, 261	187, 248, 444	146, 248, 444	146, 248, 444	243, 257, 274	257, 274	148, 274	148, 274	159, 252, 278	243, 252, 278	248, 444	-
AFS006	258, 308	-	-	-	308	300	308	-	-	-	-	-
AFS145	210, 229	194, 210	194, 210	196, 210	210, 229	200, 219	-	-	-	-	196, 210	196, 210
AFRA03F03	248, 373, 400, 455	150, 287, 355, 391, 418, 473	287	152, 196, 287, 391	243, 274, 400, 464	235, 243, 274, 400, 455	-	-	-	-	-	-
AFA01A08	135, 308	108, 135, 308	122	129, 133, 420, 428	261, 271, 282	133, 149, 308	-	-	-	-	-	-
AFB02C10	310	-	-	-	-	-	-	-	-	-	-	-
AFAA03F01	236, 254	308, 408	408	323, 408	232, 250	239, 254	271	-	-	-	308	-
AFA03G09	291	-	-	-	-	291	-	-	-	-	-	-
AFB02G04	162, 166, 174, 177	-	-	-	174, 177	162, 174	162	162	-	-	-	-
AFA07G06	139, 170, 185	-	-	-	214	135, 139, 148, 172,	185, 191	-	-	148, 185	-	163, 198

ACC008	721	177, 292, 547, 579, 753	177, 292, 516, 547, 611, 674, 689, 753	516, 547, 579, 674, 753	-	-	-	-	292, 737	737	674, 737	-
ACE010-1	174	187	189	187	176, 222	213	183	183	178	178	183, 187	-
ACE127	248	240, 260	240, 276	240, 260, 276	212, 244	212, 244	224	224	252	252	236, 252, 264	252
ACM071	158, 163	160, 161	161	158, 161	-	-	-	-	-	-	-	-
AFAT05E04	193, 282	193, 223	191, 218, 232, 245	193	209, 291, 355	193, 209	191	191	-	198	193, 236	193, 227
AFAT05G01	158, 164, 182, 189, 310, 340	-	-	-	156, 189, 219, 281, 295	156, 295	-	-	-	-	-	-
AFRT01H05	-	407	393	407	281, 393	281	281	-	281	281	-	281

^z The SSR loci were described by Tsukazaki et al. (2007) and Kuhl et al. (2004).

^y “-” indicates absence of amplified fragments in the species.

^x intraspecific hybrid between shallot and bulb onion