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Role of Alpha-actinin-4 in the Regulation of Endocytosis

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Abstract Alpha-actinin-4, originally identified as an actin-binding protein, has been associated with cell motility and cancer invasion. However, it forms complexes with a diverse array of partner proteins and is speculated to exert several distinct functions based on the characteristics of these proteins. Immunoprecipitation and mass spectrometric analysis reveal that alpha-actinin-4 forms native complexes with several partner proteins in 22RV1 cells, including β/γ -actin, clathrin heavy chain and nonmuscular myosin heavy chain. Clathrin is a coat protein that covers the internalized membrane pit and plays a central role in early endocytosis. We identified other clathrin-related and unrelated cargo proteins, including dynamin, adaptin-γ, β-NAP and p47A, that interact with alpha-actinin-4. Immunofluorescence microscopy revealed that dynamin and clathrin are co-localized with alpha-actinin-4 at the dorsal sites of membrane rufflings, and the transfection of ACTN4 cDNA facilitates the transport of transferrin into the peri-nuclear endosomes. In this review, we report that alpha-actinin-4 is involved in the regulation of endocytosis. Endocytosis enforces termination of signalling evoked by the cell surface receptors and regulates recycling of receptors and ligands. The change of alpha-actinin-4 expression levels may therefore induce aberrations in the intracellular trafficking of various cell surface molecules, such as growth factors and receptors.

Key words: actinin-4, endocytosis, clathrin, proteomics

Introduction

Alpha-actinin-4, originally identified as an actin-binding protein, has been associated with cell motility and cancer invasion. However, it forms complexes with a diverse array of partner proteins and is speculated to exert several distinct functions based on the characteristics of these proteins. In this review article, we report that alpha-actinin-4 acts as a regulator of endocytosis.

Partner proteins of alpha-actinin-4

Alpha-actinin-4 interacts with a diverse array of partner proteins (Table 1). Immuno-

precipitation of native alpha-actinin-4-containing complex using anti-alpha-actinin-4 antibody identified proteins that interact with alpha-actinin-4.⁵⁾ This is known to prevent the identification of artificial interactions, such as those commonly observed in transfection experiments.²⁾⁶⁾ Immunoprecipitation of the human prostate cancer 22RV1 cell lysate with anti-alpha-actinin-4 antibody revealed that several proteins selectively communoprecipitated with alpha-actinin-4. The proteins identified by mass spectrometry and a database search ⁷⁻¹⁰⁾ and confirmed by Western blot analysis ¹¹⁾¹²⁾ revealed that myosin heavy chain nonmuscle form A (gi:625305),

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Table 1 Partner proteins of alpha-actinin-4

Partner	Function
Actin (ACTB) ¹⁾	Scaffolding protein
Alpha chain of type XVII collagen/BP180 (COL17A1) ^{24) 25)}	Alpha chain of type XVII collagen.
Beta-catenin (CTNNB1) ⁶⁾	Adherens junction protein
PDZ-LIM protein Elfin/CLP-36 (PDLIM1) ²⁶⁾	Actin stress fiber formation and focal adhesion
Membrane associated guanylate kinase (MAGI-1) ²⁷⁾	Scaffolding protein at cell-cell junctions
Densin-180 (LRRC7) ^{28) 29)}	Transmembrane synaptic adhesion molecule
Zonula occludens-1 (TJP1) ³⁰⁾	Tight junction protein 1
Androgen receptor (NR3C1) ¹⁸⁾	Transcription factor
	Regulator of other transcription factors
Intercellular adhesion molecule 1 (ICAM1) ³¹⁾	Rhinovirus receptor
Aquaporin-2 (AQP2) ³²⁾	Water channel protein located
Na(+)/H(+) exchanger (NHE)3 kinase A regula-	Sodium/hydrogen exchanger
tory protein (E3KARP) ^{33) 34)}	
CART complex (hrs/Actn4/brain expressed ring	Organelle transport
finger protein (BERP)/myosin V) ^{16) 35)}	Neurite outgrowth
Junctional Rab13-binding protein (JRAB)/ mole-	Endocytic recycling
cule interacting with CasL-like 2 (MICALL2) ¹⁷⁾	
Endothelial nitric oxide synthase (NOS3) ³⁶⁾	Nitric oxide synthase
Inducible nitric oxide synthase (NOS2A) ³⁷⁾	Nitric oxide synthase
DNaseY (DNASE1L3) ³⁸⁾	DNase family
p65/RelA subunit of NF-kappaB (RELA) ³⁹⁾⁴⁰⁾	NFκB pathways
PDZ and LIM domain 2 (PDLIM2) ⁴¹⁾	NFκB pathways
Histone deacetylase 7 (HDAC7) ⁴²⁾	Transcriptional regulation
	Cell cycle progression
HLA-B associated transcript 1 (BAT-1) ⁴³⁾	Splicing factor
Rac serine/threonine protein kinase (AKT1) ⁴⁴⁾	Mediator of growth factor-induced
	neuronal survival
Glucose transporter 4 (GLUT4) ⁴⁵⁾	Insulin-regulated facilitative glucose transporter
OK/SW-CL.16 protein (OK/SW-CL.16) ⁴⁶⁾	Signaling proteins in podocytes
RN-tre (USP6NL) ⁴⁷⁾	Rab5 GTPase-activating protein
Plasminogen activator inhibitor type 1 (SERPINE1) ⁴⁸⁾	Plasminogen activator inhibitor
Nuclear transcription factor Y (NF-Y) ⁴⁹⁾	CCAAT-binding factor

clathrin heavy chain 1 (gi:4758012), tubulin- α 2/ β -1 (gi:135448 and gi:34740335) and cytoskeletal actin- γ 1 (gi: 4501887) co-immunoprecipitate with alpha-actinin-4.

Clathrin and endocytosis

Endocytosis is characterized by internalization of molecules from the cell surface into internal membrane compartments. Clathrin is a coat protein that covers the internalized membrane pit and plays a central role in early endocytosis.¹³⁾ The internalization of various cargo proteins and lipids from the mammalian cell surface membrane occurs through the clathrin and lipid-raft endocytic pathways, which are closely related with exocytosis, phagocytosis and pinocytosis. Clathrin triskelia assembles into a polygonal lattice at the plasma membrane to form coated pits that bud and pinch off from the

membrane in a dynamin-dependent manner and give rise to clathrin-coated vesicles in clathrin dependent endocytosis. Moreover, clathrin-binding adaptors bind to clathrin directly to initiate this process and bind to cargo proteins, thus mediating their endocytosis. Clathrin-coated vesicles are uncoated following endocytosis and fuse with the early endosome. The early endosome is therefore a key control point for sorting receptors, which can be directed to recycling endosomes back to the cell surface, or directed to the intralumenal vesicle of multivesicular endosomes, including late endosome and lysosome for degradation.

Association of endocytosis-related proteins and alpha-actinin-4

Analysis of the immunoprecipitates identified with anti-alpha-actinin-4 antibody and western blotting using antibodies against other endocytosis-related proteins revealed that Adaptin-δ, β-NAP, p47A and dynamin were co-immunoprecipitated with anti-alphaactinin-4 antibody. Confocal immunofluorescence microscopy revealed that the clathrin heavy chain and alpha-actinin-4 proteins partially co-localize at the dorsal sites of membrane ruffling, and that dynamin and alphaactinin-4 co-localize at the membrane ruffling and protrusions. Dynamin assembles around the necks of clathrin-coated pits and assists in pinching of the vesicles from the plasma membrane.¹³⁾ Adaptin-δ, β-NAP and p47A were identified as the main components of the adaptor-related protein complex 3,14) which appears to function independently of clathrin.¹⁵⁾

Regulation of alpha-actinin-4 for endocytosis

Alpha-actinin-4 interacts with various endocytosis-related proteins.⁵⁾ We confirmed the role of alpha-actinin-4 regulation in endocytosis. The expression of HA-ACTN4 in HeLa cells induced the transport of fluorescence-labelled transferrin to peri-nuclear endosomes, confirming the functional involvement of alpha-actinin-4 in endocytosis. Similarly the alpha-actinin-4-Hrs-BERP-myosin V complex was reported to be associated with transferrin receptor recycling.¹⁶⁾ Alpha-actinin-4 is known to associate with JRAB/MICAL-L2, which mediates the endocytic recycling of

occludin.¹⁷⁾ Alpha-actinin-4 is also known to interact closely with the androgen receptor, clathrin heavy chain and serine-threonine kinase C delta.¹⁸⁾ Endocytosisregulating GTPase proteins Rab5 has been observed to interact with alpha-actinin-4.¹⁹⁾ ACTN4 was identified as one of genes upregulated by Rac1 and Cdc42 by cDNA microarray analysis.²⁰⁾ Cdc42 is necessary for actin polymerization during compensatory endocytosis.²¹⁾ Moreover, alpha-actinin-4 was preferentially concentrated into circular ruffling and macropinocytosis of mouse macrophages.²²⁾ Furthermore, alpha-actinin-4 expression at the apical surface of the intestinal tract epithelium suggests its involvement in the pinocytic absorption of nutrition.²³⁾

Conclusions

Alpha-actinin-4 interacts with a diverse array of partner proteins, suggesting a multitude of functions based on the specific interactions. We have demonstrated that alpha-actinin-4 interacts with various endocytosis-related proteins. (5) Changes in alpha-actinin-4 expression levels may induce aberrations in intracellular trafficking of various cell surface molecules, such as growth factors and receptors, thereby affecting cell signalling processes.

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